

Introduction to RBM package

Dongmei Li

October 30, 2025

Clinical and Translational Science Institute, University of Rochester School of Medicine and Dentistry, Rochester, NY 14642-0708

Contents

1 Overview	1
2 Getting started	2
3 RBM_T and RBM_F functions	2
4 Ovarian cancer methylation example using the RBM_T function	6

1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code. Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
<code>ordfit_t</code>	1000	-none-	numeric
<code>ordfit_pvalue</code>	1000	-none-	numeric
<code>ordfit_beta0</code>	1000	-none-	numeric
<code>ordfit_beta1</code>	1000	-none-	numeric
<code>permutation_p</code>	1000	-none-	numeric
<code>bootstrap_p</code>	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 20

> which(myresult$permutation_p<=0.05)

[1] 61 69 87 96 219 249 365 486 495 496 505 512 565 627 678 720 743 756 832
[20] 841

> sum(myresult$bootstrap_p<=0.05)

[1] 8

> which(myresult$bootstrap_p<=0.05)

[1] 4 273 360 363 505 527 681 887

> permutation_adjp <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adjp<=0.05)

[1] 3

> bootstrap_adjp <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adjp<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 7

> which(myresult2$bootstrap_p<=0.05)

[1] 20 97 435 551 590 698 827

> bootstrap2_adjp <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adjp<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000  -none-  numeric
ordfit_pvalue 3000  -none-  numeric
ordfit_beta1  3000  -none-  numeric
permutation_p 3000  -none-  numeric
bootstrap_p   3000  -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 70

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 61

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 62

> which(myresult_F$permutation_p[, 1]<=0.05)

 [1]  7 12 35 45 46 67 105 112 133 156 171 205 209 230 252 262 263 277 285
[20] 289 313 319 327 331 338 399 411 426 429 433 446 450 463 472 475 488 495 499
[39] 524 539 540 566 579 623 631 636 648 674 675 705 728 761 769 770 785 826 834
[58] 838 840 862 887 890 909 917 946 947 963 984 985 989

> which(myresult_F$permutation_p[, 2]<=0.05)

 [1]  7 12 35 40 45 46 67 105 112 133 171 209 230 263 277 285 289 313 319
[20] 325 331 338 339 399 401 411 426 429 433 463 472 475 488 499 524 566 579 623
[39] 631 636 648 652 705 728 758 761 769 793 826 834 838 840 862 887 909 917 946
[58] 947 963 985 989

> which(myresult_F$permutation_p[, 3]<=0.05)

 [1]  7 12 45 46 67 105 112 156 171 209 230 262 263 277 285 289 313 319 331
[20] 338 399 420 426 433 450 472 475 488 499 524 539 540 566 579 624 631 636 648
[39] 674 684 695 705 728 761 769 770 793 825 826 834 838 840 862 887 909 917 946
[58] 947 963 974 985 989

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 14

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 12

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 16

> which(con2_adjp<=0.05/3)

[1] 46 105 263 289 426 631 648 728 826 840 887 947

> which(con3_adjp<=0.05/3)

[1] 45 46 105 263 426 566 631 636 648 705 728 826 840 887 947 963

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p   3000   -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 67

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 58

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 63

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 5 41 42 45 50 51 56 57 63 65 72 103 109 114 135
[16] 153 171 212 213 216 227 273 276 287 289 294 354 384 393 399
[31] 417 425 426 431 435 439 454 473 491 541 550 556 559 586 607
[46] 636 653 694 705 725 737 760 764 792 820 822 826 830 849 853
[61] 866 899 939 957 981 996 1000

```

```
> which(myresult2_F$bootstrap_p[, 2]<=0.05)
```

```

[1] 5 19 41 50 56 57 63 65 72 103 109 114 135 147 153
[16] 171 216 227 273 287 289 338 384 393 399 417 426 431 436 439
[31] 454 473 491 529 541 556 559 586 636 653 669 671 694 705 731
[46] 737 764 822 826 830 849 861 866 899 939 960 981 1000

```

```
> which(myresult2_F$bootstrap_p[, 3]<=0.05)
```

```

[1] 5 19 41 42 45 50 51 56 57 63 65 72 103 114 116
[16] 135 153 171 212 213 216 227 273 287 289 325 393 399 426 431
[31] 439 473 491 504 529 541 556 559 586 636 668 669 671 694 705
[46] 737 760 764 792 820 822 826 830 849 861 866 899 939 943 957
[61] 981 996 1000

```

```
> con21_adjp <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
```

```
> sum(con21_adjp<=0.05/3)
```

```
[1] 13
```

```
> con22_adjp <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
```

```
> sum(con22_adjp<=0.05/3)
```

```
[1] 5
```

```
> con23_adjp <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
```

```
> sum(con23_adjp<=0.05/3)
```

```
[1] 8
```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following

codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "/tmp/RtmpH9xrEa/Rinst277e277b870136/RBM/data"
```

```
> data(ovarian_cancer_methylation)
```

```
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59031	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	
exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		
exmdata8[, 2]			
Min. :0.01357			
1st Qu.:0.04387			
Median :0.09282			
Mean :0.28679			
3rd Qu.:0.57217			
Max. :0.96268			

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
```

```
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
```

```
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
```

```
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 47

> sum(diff_results$permutation_p<=0.05)

[1] 53

> sum(diff_results$bootstrap_p<=0.05)

[1] 62

> ordfit_adj <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj<=0.05)

[1] 0

> perm_adj <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj<=0.05)

[1] 2

> boot_adj <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj<=0.05)

[1] 9

> diff_list_perm <- which(perm_adj<=0.05)
> diff_list_boot <- which(boot_adj<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
19 cg00016968 0.8062848             NA      0.8144082      0.8362318
764 cg00730260 0.9047127      0.9054229      0.9100268      0.9125861
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
19      0.8083138      0.7330644      0.8296834      0.8491780
764      0.9057589      0.8876047      0.9075630      0.9094679
      diff_results$ordfit_t[diff_list_perm]
19                                 -2.547097
764                                -1.560713
      diff_results$permutation_p[diff_list_perm]
19                                 0
764                                 0

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)

```


	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
16	cg00014085	0.05906804	0.04518973	0.04211710	0.03665208
106	cg00095674	0.07076291	0.05045181	0.03861991	0.03337576
131	cg00121904	0.15449580	0.17949750	0.23608110	0.24354150
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
346	cg00331237	0.05972383	NA	0.08204769	0.08345662
522	cg00503840	0.16919790	0.21350810	0.23958290	0.24448410
772	cg00743372	0.03922780	0.02919634	0.02187972	0.02568053
887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
16	0.04222944	0.05324246	0.03728026	0.04062589	
106	0.04693030	0.06837343	0.04534005	0.03709488	
131	0.17352980	0.12564280	0.18193170	0.20847670	
146	0.67191510	0.63137380	0.47929610	0.45428300	
346	0.05372019	0.06241126	0.06955040	0.09140985	
522	0.25154270	0.16469600	0.23368520	0.27977040	
772	0.02796053	0.03512214	0.02575992	0.02093909	
887	0.50259740	0.40111730	0.56646700	0.54552980	
911	0.08633986	0.06765189	0.09070268	0.12417730	
	diff_results\$ordfit_t[diff_list_boot]				
16	1.954876				
106	2.887876				
131	-3.562745				
146	5.636263				
346	-3.328798				
522	-2.442169				
772	1.885560				
887	-3.368752				
911	-3.490240				
	diff_results\$bootstrap_p[diff_list_boot]				
16	0				
106	0				
131	0				
146	0				
346	0				
522	0				
772	0				
887	0				
911	0				