

# Package ‘ENmix’

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BeadChip

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---

B2M

*Converting methylation beta value to M value.*

---

### Description

Convert methylation beta value to M value.

### Usage

B2M(x)

**Arguments**

x                      An numeric matrix with values between 0 and 1

**Details**

Methylation beta value is calculated as  $\beta = M/(M+U+a)$ . M is methylated intensity, U is unmethylated intensity, and a is a constant offset (by default,  $a=100$ ). M value is calculated as  $M = \log_2((M+a)/(U+a))$ . M or U is usually greater than 1000, so a is negligible for most probes. if  $a=0$ , then  $M = \log_2(\beta)/(1-\beta)$ .

**Value**

A matrix of M values

**Author(s)**

Zongli Xu

**Examples**

```
if (require(minfiData)){
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  mdat = getmeth(rgSet)
  beta = getB(mdat, "Illumina")
  m = B2M(beta)
}
```

---

calcdetP

*To calculate detection P values*

---

**Description**

Calculation of detection P values based on negative internal control probes or out of the band (oob) probes

**Usage**

```
calcdetP(rgSet, detPtype = "negative")
```

**Arguments**

rgSet                      An object of class rgDataSet  
 detPtype                      Calculation of detection P values based on negative internal control ("negative") probes or out of the band ("oob") probes

**Value**

An numerical matrix of detection P values, with row for CpGs and column for samples

**Author(s)**

Zongli Xu

## References

Wanding Zhou et al. SeSAMe: reducing artifactual detection of DNA methylation by Infinium BeadChips in genomic deletions, *Nucleic Acids Research*, 2018

## Examples

```
path <- file.path(find.package("minfiData"), "extdata")
rgSet <- readidat(path = path, recursive = TRUE)
detp=calcdetP(rgSet, detPtype = "negative")
detp2=calcdetP(rgSet, detPtype = "oob")
```

---

combp

*Identification of differentially methylated regions*

---

## Description

To identify differentially methylated regions using a modified comb-p method

## Usage

```
combp(data, dist.cutoff=1000, bin.size=310, seed=0.01,
      region_plot=TRUE, mht_plot=TRUE, nCores=10, verbose=TRUE)
```

## Arguments

data	A data frame with colname name "chr", "start", "end", "p" and "probe", indicating chromosome (1,2,3,...,X,Y), chromosome start and end position, P value and probe names
dist.cutoff	Maximum distance in base pair to combine adjacent DMRs
bin.size	bin size for autocorrelation calculation
seed	FDR significance threshold for initial selection of DMR region
region_plot	If TRUE, regional plots will be generated
mht_plot	If TRUE, mahattan plot will be generated
nCores	Number of computer cores will be used in calculation
verbose	If TRUE, detailed running information will be printed

## Details

The input should be a data frame with column names "chr", "start", "end", "p", and "probe", indicating chromosome number, start position, end position, P value and probe name. The function use a modified comb-p method to identify differentially methylated regions. DMR results will be stored in a file with name `resu_combp.csv`. If plot options were selected, two figure files will be generated: `mht.jpg` and `region_plot.pdf`.

## Author(s)

Liang Niu, Zongli Xu

## References

Pedersen BS1, Schwartz DA, Yang IV, Kechris KJ. Comb-p: software for combining, analyzing, grouping and correcting spatially correlated P-values. *Bioinformatics* 2012

Zongli Xu, Changchun Xie, Jack A. Taylor, Liang Niu, ipDMR: Identification of differentially methyl-ated regions with interval p-values, *Bioinformatics* 2020

## Examples

```
dat=simubed()
names(dat)
#seed=0.1 is only for demonstration purpose, it should be smaller than 0.05 or 0.01 in actual study.
combp(data=dat,seed=0.1)
```

---

ctrlsva	<i>Non-negative internal control surrogate variables</i>
---------	--

---

## Description

Surrogate variables derived based on intensity data for non-negative internal control probes.

## Usage

```
ctrlsva(rgSet,percvar=0.95,npc=1,flag=1)
```

## Arguments

rgSet	An object of class rgDataSet or RGChannelSet.
percvar	Minimum percentage of data variations can be explained by surrogate variables, range from 0 to 1,default is 0.95
npc	Number of surrogate variables, default is 1
flag	1: select number of surrogate variables based on argument percvar; 2: select number of surrogate variables based on argument npc

## Value

The function will return an numerical matrix with columns indicating surrogate variables and rows corresponding to samples. These variables can be used in association analysis to adjust for experimental batch effects.

## Author(s)

Zongli Xu

## References

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. *Nucleic Acids Research* 2015.

**Examples**

```

if (require(minfiData)) {
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  sva <- ctrlsva(rgSet)
}

```

dupicc

*Evaluation of measurement reliability using duplicate samples***Description**

The function can be used to evaluate duplicate samples by calculating: 1) centered/un-centered Pearson's correlation coefficient between duplicates; 2) absolute difference between duplicates; 3) ICC for each CpG probes using oneway or twoway model.

**Usage**

```

dupicc(dat, dupid, mvalue=FALSE, center=TRUE, nCores=2, qcflag=FALSE, qc=NULL,
       detPthre=0.05, nbthre=3, skipicc=FALSE, corfig=FALSE, model="oneway")

```

**Arguments**

dat	Methylation beta value matrix
dupid	A data frame with two variables, id1 and id2. The two ids in each row indicate a duplicate pair. These ids should be the same with column names of the input methylation matrix
mvalue	If TRUE, the beta value will be converted to M value for calculation of ICC
center	If TRUE, the methylation beta values will be centered for each CpG before calculation of ICC or correlation
nCores	Number of cores will be used for calculation of ICC
qcflag	Whether to perform QC before calculation of ICC
qc	QC object from function QCinfo
detPthre	If qcflag=TRUE, the methylation values with detection P value higher than the threshold will be removed before calculation
nbthre	If qcflag=TRUE, the methylation values with number of bead smaller than the threshold will be removed
skipicc	If TRUE, ICC calculation will be skipped
corfig	If TRUE, a figure will be generated to demonstrate correlations within duplicates or within non-duplicates
model	Using "oneway" or "twoway" model to calculate ICC

**Value**

icc: a data frame containing ICC and P values for each probe

dupcor: a data frame containing Pearson's correlation and averaged absolute difference between duplicates.

**Author(s)**

Zongli Xu

**References**

Zongli Xu, Jack A Taylor. *Reliability of DNA methylation measures using Illumina methylation BeadChip*. Epigenetics 2020

**Examples**

```
if (require(minfiData)){
  path <- file.path(find.package("minfiData"),"extdata")
  rgSet <- readidat(path = path,recursive = TRUE)
  mdat=getmeth(rgSet)
  beta=getB(mdat,"Illumina")
  dupidx=data.frame(id1=c("5723646052_R02C02","5723646052_R04C01","5723646052_R05C02"),
  id2=c("5723646053_R04C02","5723646053_R05C02","5723646053_R06C02"))
  iccresu<-dupicc(dat=beta,dupid=dupidx)
}
```

estimateCellProp

*Cell type proportion estimator***Description**

To estimates relative proportion of underlying cell types in a sample based on reference methylation data of pure cell types.

**Usage**

```
estimateCellProp(userdata,refdata="FlowSorted.Blood.450k",
  cellTypes=NULL,nonnegative = TRUE,nProbes=50,
  normalize=TRUE,refplot=FALSE)
```

**Arguments**

userdata	The input can be rgDataSet,methDataSet, MethylSet,RGChannelSet or methylation beta value matrix.
refdata	Reference data set will used. Current option: "FlowSorted.Blood.450k", "FlowSorted.DLPFC.450k", "FlowSorted.CordBlood.450k", "FlowSorted.CordBloodCombined.450k", "FlowSorted.CordBloodNorway.450k" or "FlowSorted.Blood.EPIC".
cellTypes	Specific set of cell type data in reference data will be used for deconvolution. if "NULL" all cell types data will be used. see details for possible cell types
normalize	TRUE or FALSE, if TRUE, quantile normalization on methylated and unmethylated intensities will be performed.
nonnegative	TRUE or FALSE. If TRUE, the estimated proportions will be constrained to nonnegative values
nProbes	Number of best probes for each cell types will be used for the estimation.
refplot	TRUE or FALSE. IF TRUE, refdata distribution and heatmap will be plotted for inspection of reference dataset.

## Details

This function use the method of Houseman et al (2012) to estimate cell type proportions based on reference DNA methylation data.

The following reference datasets can be used to assist the estimation. User should select a reference most resemble to user's data in tissue, age, and array type.

FlowSorted.Blood.450k: consisting of 450K methylation data for 60 blood samples from 6 male adults. Six samples for each of the cell types: Bcell CD4T CD8T Eos Gran Mono Neu NK PBMC WBC; See Reinius et al. 2012 for details.

FlowSorted.CordBlood.450k: consisting of 450k methylation data for 104 cord blood samples from 17 male and female individuals. Cell type (# samples) are: Bcell(15) CD4T(15) CD8T(14) Gran(12) Mono(15) NK(14) nRBC(4) WholeBlood(15). See Bakulski et al. Epigenetics 2016 for details.

FlowSorted.CordBloodNorway.450k: consisting of 450K methylation data for 77 cord blood samples from 11 individuals (6 girls and 5 boys). 11 samples for each of the cell types: Bcell CD4T CD8T Gran Mono NK WBC. See P Yousefi et al Environ. Mol. Mutagen 2015 for details.

FlowSorted.Blood.EPIC: consisting of EPIC methylation data for 37 magnetically sorted blood cell references from 12 individuals. See LA Salas et al. 2018 for details.

FlowSorted.DLPFC.450k: consisting of 450K methylation data for 58 brain tissue samples from 29 individuals. 15 females and 14 males, 6 Africans and 23 Caucasians, age range from 13 to 79. 29 samples for each of the cell types: NeuN\_neg and NeuN\_pos. See Guintivano et al. 2013 for details.

FlowSorted.CordBloodCombined.450k: consisting of 289 combined umbilical cord blood cells samples assayed by Bakulski et al, Gervin et al., de Goede et al., and Lin et al. see <https://github.com/immunomethylomic/FlowSorted.CordBloodCombined.450k>. for details.

## Value

The output is a data frame composed of the estimates of cell type proportions with columns indicate cell types and rows indicate samples.

## Author(s)

Zongli Xu

## References

EA Houseman, WP Accomando, DC Koestler, BC Christensen, CJ Marsit, HH Nelson, JK Wiencke and KT Kelsey. *DNA methylation arrays as surrogate measures of cell mixture distribution*. BMC bioinformatics (2012) 13:86.

## Examples

```
require(minfiData)
path <- file.path(find.package("minfiData"), "extdata")
#based on rgDataset
rgSet <- readidat(path = path, recursive = TRUE)
celltype=estimateCellProp(userdata=rgSet, refdata="FlowSorted.Blood.450k",
                          nonnegative = TRUE, normalize=TRUE)
#using methDataSet
qc=QCinfo(rgSet)
mdat<-preprocessENmix(rgSet, bgParaEst="oob", dyeCorr="RELIC",
                      QCinfo=qc, nCores=6)
```

```

celltype=estimateCellProp(userdata=mdat,refdata="FlowSorted.Blood.450k",
                          nonnegative = TRUE,normalize=TRUE)
mdat<-norm.quantile(mdat, method="quantile1")
#using beta value
beta<-rcp(mdat,qcscore=qc)
celltype=estimateCellProp(userdata=beta,refdata="FlowSorted.Blood.450k",
                          nonnegative = TRUE)

```

---

freqpoly

*Frequency polygon plot*


---

### Description

Similar to histogram, frequency polygon plot can be used to inspect data distribution.

### Usage

```

freqpoly(mat, nbreaks=15, col="black", xlab="", ylab="Frequency",
         type="l", append=FALSE, ...)

```

### Arguments

mat	A numeric vector
nbreaks	Number of bins for frequency counting
col	color code
xlab	x-axis lable
ylab	y-axis lable
type	character indicating the type of plotting; actually any of the 'type's as in 'plot.default'.
append	TRUE or FALSE, whether to create a new figure or append to the current figure.
...	Further arguments that get passed to the function "plot"

### Value

Frequency polygon plot.

### Author(s)

Zongli Xu

### References

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015.

### Examples

```

freqpoly(rnorm(1000))

```

---

getB *Extract methylation Beta values.*

---

### Description

Extract Methylation Beta value,  $\text{Beta} = \text{Meth} / (\text{Meth} + \text{Unmeth} + \text{offset})$

### Usage

```
getB(mdat, type="Illumina", offset=100)
```

### Arguments

mdat	An object of class methDataSet or MethylSet.
type	type="Illumina" sets offset=100 as per Genome Studio software.
offset	Regularization factor in calculating beta ratio, 100 in default

### Value

Methylation Beta value =  $\text{Meth} / (\text{Meth} + \text{Unmeth} + \text{offset})$ . Meth is methylated intensity matrix, Unmeth is unmethylated intensity matrix.

### Author(s)

Zongli Xu

### Examples

```
if (require(minfiData)){
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  mdat=getmeth(rgSet)
  beta=getB(mdat, "Illumina")
}
```

---

getCGInfo *CpG probe annotation information*

---

### Description

Extract CpG probe annotation information from an rgDataSet

### Usage

```
getCGInfo(rgSet, type="IandII")
```

### Arguments

rgSet	An object of class rgDataSet
type	One of the following options "I", "II", "IandII", "ctrl", indicating type I, type II type I & II or control probes type

**Value**

An object of data frame class

**Author(s)**

Zongli Xu

**Examples**

```
require(minfiData)
path <- file.path(find.package("minfiData"), "extdata")
#based on rgDataset
rgSet <- readidat(path = path, recursive = TRUE)
cginfo=getCGinfo(rgSet, type="IandII")
```

---

getmeth

*Create a methDataSet*

---

**Description**

To create a methDataSet based on a rgDataset

**Usage**

```
getmeth(rgSet)
```

**Arguments**

rgSet            An object of class rgDataSet

**Details**

CpG annotation information is incorporated in the output methDataSet object, and can be extracted using command `rowData()`.

**Value**

An object of class methDataSet

**Author(s)**

Zongli Xu

## Examples

```
require(minfiData)
path <- file.path(find.package("minfiData"), "extdata")
#based on rgDataset
rgSet <- readidat(path = path, recursive = TRUE)
meth=getmeth(rgSet)
meth
cginfo=rowData(meth)
```

---

ipdmr

*Differentially methylated regions*


---

## Description

To identify differentially methylated regions using an interval P value method

## Usage

```
ipdmr(data, include.all.sig.sites=TRUE, dist.cutoff=1000, bin.size=50,
       seed=0.05, region_plot=TRUE, mht_plot=TRUE, verbose=TRUE)
```

## Arguments

data	A data frame with colname name "chr", "start", "end", "p" and "probe", indicating chromosome (1,2,3,...,X,Y), chromosome start and end position, P value and probe names
include.all.sig.sites	Whether to use CpG singletons in calculation of FDR
dist.cutoff	Maximum distance in base pair to combine adjacent DMRs, and the maximum distance between CpGs where auto-correlation will be calculated
bin.size	bin size for autocorrelation calculation
seed	FDR threshold for initial selection of DMR regions
region_plot	If TRUE, regional plots will be produced for each DMR
mht_plot	If TRUE, a p-value mahattan plot with marked DMRs will be produced
verbose	Whether to output detailed information

## Details

The input should be a data frame with column names "chr", "start", "end", "p" and "probe", indicating chromosome, start and end position, P value and probe name. The function will use a novel interval p value method to identify differentially methylated regions. DMR results will be stored in a file with name resu\_ipdmr.csv. If plot options were selected, two figure files will be generated: mht.jpg and region\_plot.pdf.

## Author(s)

Liang Niu, Zongli Xu

## References

Zongli Xu, Changchun Xie, Jack A. Taylor, Liang Niu, ipDMR: Identification of differentially methylated regions with interval p-values, Bioinformatics 2020

## Examples

```
dat=simubed()
names(dat)
#seed=0.1 is only for demonstration purpose, it should be smaller than 0.05 or 0.01 in actual study.
ipdmr(data=dat,seed=0.1) #seed=0.1
```

---

M2B

---

*Converting methylation M value to beta value.*


---

## Description

Converting methylation M value to methylation beta value.

## Usage

```
M2B(x)
```

## Arguments

x                      An numeric matrix

## Details

Methylation beta value is calculated as  $\beta = M / (M + U + a)$ . M is methylated intensity, U is unmethylated intensity, and a is a constant offset (by default,  $a = 100$ ). M value is calculated as  $M = \log_2((M + a) / (U + a))$ . M or U is usually greater than 1000, so a is negligible for most probes. if  $a = 0$ , then  $\beta = 2^M / (2^M + 1)$ .

## Value

A matrix of methylation Beta values.

## Author(s)

Zongli Xu

## Examples

```
if (require(minfiData)){
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  mdat = getmeth(rgSet)
  beta = getB(mdat, "Illumina")
  m = B2M(beta)
  b = M2B(m)
}
```

---

methDataSet-class      *Class "methDataSet"*

---

### Description

A class for storing Illumina methylation array methylated and unmethylated intensity data, and CpG annotation information.

### Usage

```
methDataSet(Meth = new("matrix"), Unmeth = new("matrix"),
            rowData=new("DataFrame"),...)
```

### Arguments

Meth	A matrix of methylated intensity values with row for CpGs and column for samples
Unmeth	A matrix of unmethylated intensity values with row for CpGs and column for samples
rowData	A dataframe contains CpG annotation information
...	Other arguments for class SummarizedExperiment

### Details

CpG annotation information is incorporated in the object, and can be extracted using command `rowData`

### Value

An object of class `methDataSet`

### Examples

```
showClass("methDataSet")
```

---

methscore      *DNA Methylation predictors*

---

### Description

To calculate various methylation predictors, including DNA methylation age, plasma protein levels, exposures etc.

### Usage

```
methscore(datMeth, datPheno=NULL, fastImputation=FALSE, normalize=TRUE,
          GrimAgeComponent=3, AgeResid=FALSE)
```

**Arguments**

datMeth	Methylation beta value matrix with CpG names(row names) and sample ids(column names).
datPheno	Phenotype data, must include columns SampleID, Age (year) and Female (0:male,1:female). Age and Female information are required to calculate DNAmFitAge and PC-Clocks
fastImputation	Logical. If "TRUE" mean methylation values will used for imputation, if "FALSE", KNN nearest neighbor method will be used.
normalize	Logical. If TRUE, user data will be normalized to a reference data using a modified RCP method
GrimAgeComponent	Which version of GrimAgeComponent will be used to calculate bAge and DNAm-FitAge. 1: GrimAgeV1; 2: GrimAgeV2 and 3: PCGrimAge.
AgeResid	Logical. If TRUE, age residuals (age acceleration) variables will be calculated.

**Details**

The original publications (see references) provided reference methylation mean values for most methylation predictors, and thus these values were used directly for normalization and imputations. For the following predictors without reference value in their original publications,

PEDBE,EpiToc,EpiToc2,Zhang10CpG,Horvath2,MiAge,DNAmTL,PEDBE,GACPC, GARPC,GARRPC,Bohlin and Knight, we used Sister Study data (~5000 samples) to derive a set of reference methylatin values. If UserRef was provided, it will be used to replace the Sister Study reference values for these predictors. If set ForceUserRef=TRUE, UserRef will be used for all predictors in normalization and CpG imputation.

**Value**

A data frame with rows for samples and columns for methylation predictors. Output file "summary\_CpG\_count.csv" has information about CpG counts and reference for each predictor.

**Author(s)**

Zongli Xu

**References**

- Belsky et al. *DunedinPACE, a DNA methylation biomarker of the pace of aging*. *Elife*. 2022 Jan 14;11:e73420. doi: 10.7554/eLife.73420
- Bernabeu et al. *Refining epigenetic prediction of chronological and biological age*. *Genome Med*. 2023 Feb 28;15(1):12. doi: 10.1186/s13073-023-01161-y
- Bohlin et al. *Prediction of gestational age based on genome-wide differentially methylated regions*. *Genome Biol*. 2016 Oct 7;17(1):207. doi: 10.1186/s13059-016-1063-4.
- Hannum et al. *Genome-wide methylation profiles reveal quantitative views of human aging rates*. *Mol Cell*. 2013 Jan 24;49(2):359-367. doi: 10.1016/j.molcel.2012.10.016. Epub 2012 Nov 21.
- Higgins-Chen et al. *A computational solution for bolstering reliability of epigenetic clocks: Implications for clinical trials and longitudinal tracking*. *Nat Aging*. 2022 Jul; 2(7): 644–661.
- Horvath. *DNA methylation age of human tissues and cell types*. *Genome Biol*. 2013;14(10):R115. doi: 10.1186/gb-2013-14-10-r115.

- Horvath et al. *Epigenetic clock for skin and blood cells applied to Hutchinson Gilford Progeria Syndrome and ex vivo studies*. *Aging (Albany NY)*. 2018 Jul 26;10(7):1758-1775. doi: 10.18632/aging.101508.
- Knight et al. *An epigenetic clock for gestational age at birth based on blood methylation data*. *Genome Biol*. 2016 Oct 7;17(1):206. doi: 10.1186/s13059-016-1068-z.
- Lee et al. *Placental epigenetic clocks: estimating gestational age using placental DNA methylation levels*. *Aging (Albany NY)*. 2019 Jun 30; 11(12): 4238–4253.
- Levine et al. *An epigenetic biomarker of aging for lifespan and healthspan*. *Aging (Albany NY)*. 2018 Apr; 10(4): 573–591.
- Li et al. *Derivation and validation of an epigenetic frailty risk score in population-based cohorts of older adults*. *Nat Commun*. 2022 Sep 7;13(1):5269. doi: 10.1038/s41467-022-32893-x.
- Lu et al. *DNA methylation-based estimator of telomere length*. *Aging (Albany NY)*. 2019 Aug 31; 11(16): 5895–5923.
- McCartney et al. *Epigenetic prediction of complex traits and death*. *Genome Biol*. 2018 Sep 27;19(1):136. doi: 10.1186/s13059-018-1514-1.
- McGreevy et al. *DNAmFitAge: biological age indicator incorporating physical fitness*. *Aging(Albany NY)*. 2023 Feb 22;15(10):3904-3938. doi: 10.18632/aging.204538.
- McEwen et al. *The PedBE clock accurately estimates DNA methylation age in pediatric buccal cells*. *Proc Natl Acad Sci U S A*. 2020 Sep 22; 117(38): 23329–23335.
- Teschendorff et al. *A comparison of epigenetic mitotic-like clocks for cancer risk prediction*. *Genome Med*. 2020 Jun 24;12(1):56. doi: 10.1186/s13073-020-00752-3.
- Yang et al. *Correlation of an epigenetic mitotic clock with cancer risk*. *Genome Biol*. 2016 Oct 3;17(1):205. doi: 10.1186/s13059-016-1064-3.
- Youn et al. *The MiAge Calculator: a DNA methylation-based mitotic age calculator of human tissue types*. *Epigenetics*. 2018; 13(2): 192–206.
- Zhang et al. *Improved precision of epigenetic clock estimates across tissues and its implication for biological ageing*. *Genome Med*. 2019 Aug 23;11(1):54. doi: 10.1186/s13073-019-0667-1.
- Zhang et al. *DNA methylation signatures in peripheral blood strongly predict all-cause mortality*. *Nat Commun*. 2017 Mar 17;8:14617. doi: 10.1038/ncomms14617.
- Lu et al. *DNA methylation GrimAge strongly predicts lifespan and healthspan*. *Aging*. 2019 Jan 21;11(2):303-327. doi: 10.18632/aging.101684.
- Lu et al. *DNA methylation GrimAge version 2*. *Aging (Albany NY)*. 2022 Dec 14;14(23):9484-9549. doi: 10.18632/aging.204434. Epub 2022 Dec 14.
- Ying et al. *Causality-enriched epigenetic age uncouples damage and adaptation*. *Nat Aging*. 2024 Feb;4(2):231-246. doi: 10.1038/s43587-023-00557-0. Epub 2024 Jan 19.

## Examples

```
require(minfiData)
path <- file.path(find.package("minfiData"), "extdata")
#based on rgDataset
rgSet <- readidat(path = path, recursive = TRUE)
meth=getmeth(rgSet)
beta=getB(meth)
pheno=data.frame(SampleID=colnames(beta), Age=c(23, 45, 52, 36, 58, 43), Female=c(1, 0, 1, 1, 0, 1))
mage=methscore(datMeth=beta, datPheno=pheno)
```

---

methyAge	<i>Methylation Age estimator</i>
----------	----------------------------------

---

**Description**

To calculate Methylation Age using Hovath, Hannum or PhenoAge methods and pace of aging DunedinPACE.

**Usage**

```
methyAge(beta, fastImputation=FALSE, normalize=TRUE, nCores=2)
```

**Arguments**

beta	Methylation beta value matrix with CpG names(row names) and sample ids(column names).
fastImputation	If "TRUE" reference methylation values will be used for imputation, if "FALSE", KNN nearest neighbor method will be used.
normalize	TRUE or FALSE, if TRUE, Hovath modified BMIQ method will be used to perform normalization.
nCores	Number of cores will be used for normalization

**Value**

A data frame with rows for sample and columns for types of methylation age.

**Author(s)**

Zongli Xu

**References**

Horvath S. *DNA methylation age of human tissues and cell types*. Genome biology 2013 14:R115.

Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sada S, et al. *Genome-wide methylation profiles reveal quantitative views of human aging rates*. Molecular cell 2013 49:359-367.

Levine ME, Lu AT, Quach A, Chen BH, Assimes TL, Bandinelli S, et al. *An epigenetic biomarker of aging for lifespan and healthspan*. Aging (Albany NY) 2018 10:573-591.

Daniel W Belsky, Avshalom Caspi, David L Corcoran, et al. *DunedinPACE, a DNA methylation biomarker of the pace of aging*. eLife, 2022

**Examples**

```
require(minfiData)
path <- file.path(find.package("minfiData"), "extdata")
#based on rgDataset
rgSet <- readidat(path = path, recursive = TRUE)
meth=getmeth(rgSet)
beta=getB(meth)
mage=methyAge(beta)
```

mhtplot

*P value manhattan plot***Description**

P value manhattan plot

**Usage**

```
mhtplot(probe=NULL,chr=NULL, pos=NULL, p=NULL,color="bg",sigthre=NULL,
sigthre2=NULL,threlty=c(2,1),markprobe=NULL,markcolor="red", outf="mht",
outfmt="jpg",reducesize=0,...)
```

**Arguments**

probe	probe name
chr	Chromosome, 1,2,...,22,X,Y
pos	Chromosome positions
p	P values
color	Color scheme of manhattan plot, "bg" indicate "black and gray"
sigthre	P value of significant threshold line
sigthre2	P value of second significant threshold line
threlty	Threshold line type, default is c(2,1)
markprobe	A list of CpGs to be marked out
markcolor	Color code for marked probe, "red" in default
outf	figure file name, default "mht"
outfmt	Output figure file format, "jpg" or "eps"
reducesize	A positive interger, larger the value, smaller the eps file size. Smaller file size is achieved by skipping some densely packed data points
...	Arguments pass to plot

**Author(s)**

Zongli Xu

**Examples**

```
dat=simubed()

thre1=1E-100
dat$fdr=p.adjust(mrgd$P, method="BH")
if(sum(dat$fdr<0.05)>0){thre1=max(dat$p[dat$fdr<0.05])}

thre2=1E-7

mprobe=dat$probe[dat$p<=thre1]
mhtplot(probe=dat$probe,chr=dat$chr,pos=dat$start,p=dat$p,sigthre=thre1,sigthre2=thre2,
markprobe=mprobe,outf="mht_try",outfmt="jpg")
```

---

mpreprocess	<i>methylation data QC and preprocessing pipeline for Illuminal Bead-Chips</i>
-------------	--

---

### Description

The pipeline performs background correction, dye bias correction, inter-array normalization and probe type bias correction for HumanMethylation 450 and MethylationEPIC BeadChip data. It removes or mitigates background noise and systematic experimental bias. It also performs quality controls, identifying and excluding low quality samples and probes, removing low quality and outlier values, and performing imputation.

### Usage

```
mpreprocess(rgSet, nCores=2, bgParaEst="oob", dyeCorr="RELIC",
            qc=TRUE, qnorm=TRUE, qmethod="quantile1",
            fqfilter=FALSE, rmcr=FALSE, impute=FALSE)
```

### Arguments

rgSet	An object of class rgDataSet, methDataSet, RGChannelSetExtended, RGChannelSet or MethylSet.
nCores	Number of cores will be used for computation
bgParaEst	Method to estimate background normal distribution parameters. Possible options: "oob", "est", or "neg".
dyeCorr	Dye bias correction, "mean": correction based on averaged red/green ratio; or "RELIC": correction with RELIC method; or "none": no dye bias correction. The default is RELIC
qc	If TRUE, QC will be performed. Low quality samples and CpGs will be excluded before background correction.
qnorm	If TRUE, inter-array quantile normalization will be performed.
qmethod	Quantile normalization method. This should be one of the following strings: "quantile1", "quantile2", or "quantile3". See details in function norm.quantile.
fqfilter	If TRUE, outlier and low quality values will be filtered out.
rmcr	TRUE: excluded rows and columns with more than 5% of missing values. FALSE is in default
impute	Whether to impute missing values. If TRUE, k-nearest neighbor's methods will be used for imputation. FALSE is in default.

### Details

Function mpreprocess is a pipeline that performs methylation data preprocessing and quality controls using functions: preprocessENmix, norm.quantile, rcp, QCinfo and qcfilter. More customized preprocessing steps can be achieved using the individual functions, see user's guide.

### Value

A methylation beta value matrix with rows for CpGs and columns for samples.

**Author(s)**

Zongli Xu

**References**

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip. Nucleic Acids Research 2015.

Zongli Xu, Sabine A. S. Langie, Patrick De Boever, Jack A. Taylor1 and Liang Niu, RELIC: a novel dye-bias correction method for Illumina Methylation BeadChip, BMC Genomics, 2017

Liang Niu, Zongli Xu and Jack A. Taylor: RCP: a novel probe design bias correction method for Illumina Methylation BeadChip, Bioinformatics 2016

**See Also**

Package minfi for classes [RGChannelSet](#) and [MethylSet](#)

**Examples**

```

if (require(minfiData)) {
  #rgDataSet as input
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  beta=mpreprocess(rgSet, nCores=6, qc=TRUE, fqfilter=TRUE, rmcr=TRUE, impute=TRUE)

  #methDataSet as input
  mdat=getmeth(rgSet)
  beta=mpreprocess(mdat, nCores=6)

  #RGChannelSet as input
  beta=mpreprocess(RGsetEx, nCores=6)

  #RGChannelSetExtended as input
  sheet <- read.metharray.sheet(file.path(find.package("minfiData"), "extdata"),
    pattern = "csv$")
  rgSet <- read.metharray.exp(targets = sheet, extended = TRUE)
  beta=mpreprocess(rgSet, nCores=6, qc=TRUE, fqfilter=TRUE, rmcr=TRUE, impute=TRUE)

  #MethylSet as input
  mdat=preprocessRaw(rgSet)
  beta=mpreprocess(mdat, nCores=6)

}

```

---

multifreqpoly

*Multiple frequency polygon plot*


---

**Description**

Produce Frequency polygon plot for each column of a numeric data matrix. Similar to multidensity function, the plot can be used to inspect data distribution but with much faster speed.

**Usage**

```
multifreqpoly(mat, nbreaks=100, col=1:ncol(mat), xlab="",
              ylab="Frequency", legend = list(x = "top", fill=col,
              legend = if(is.null(colnames(mat))) paste(1:ncol(mat))
              else colnames(mat)), append=FALSE, ...)
```

**Arguments**

mat	A numeric matrix
nbreaks	The number of bins for frequency counting
col	Line plot color code, the length should be equal to the number of columns in mat
xlab	x-axis lable
ylab	y-axis lable
legend	A list of arguments that get passed to the function "legend"
append	TRUE or FALSE, whether to create a new figure or append to the current figure.
...	Further arguments that get passed to the function "plot"

**Value**

Frequency polygon plots.

**Author(s)**

Zongli Xu

**References**

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015.

**Examples**

```
x=matrix(rnorm(10000),nrow=2000,ncol=5)
multifreqpoly(x,nbreaks=15,legend=list(x="topright",fill=1:ncol(x),legend=paste("V",1:5,sep="")))

if (require(minfiData)) {
  path <- file.path(find.package("minfiData"),"extdata")
  rgSet <- readidat(path = path,recursive = TRUE)
  mraw <- getmeth(rgSet)
  beta<-getB(mraw)
  jpeg("dist_raw.jpg")
  multifreqpoly(beta,col=1:ncol(beta))
  dev.off()
}
```

---

nmode

*Estimating number of mode for each row of data*

---

## Description

Due to SNPs in CpG probe region or other unknown factors, methylation beta values for some CpGs have multimodal distribution. This function is to identify this type of probes (so called gap probes) with obvious multimodal distribution.

## Usage

```
nmode(x, minN = 3, modedist=0.2, nCores = 1)
```

## Arguments

x	A methylation beta value matrix with row for probes and column for samples.
minN	Minimum number of data points at each cluster
modedist	Minimum distance between adjacent modes
nCores	Number of cores used for computation

## Details

This function uses an empirical approach to estimate number of modes in methylation beta value for each CpG probe. By default, the function requires the distance between modes have to be greater than 0.2 in methylation beta value, and each mode clusters should have at least 3 data points or 5% of data points whichever is greater.

## Value

A vector of integers indicating number of modes. Gap probes are probes with number of modes greater than 1.

## Author(s)

Zongli Xu

## References

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015

## Examples

```
if (require(minfiData)) {  
  mdat <- preprocessRaw(RGsetEx)  
  beta=getBeta(mdat, "Illumina")  
  nmode=nmode(beta, minN = 3,modedist=0.2, nCores = 5)  
  
  path <- file.path(find.package("minfiData"),"extdata")  
  rgSet <- readidat(path = path,recursive = TRUE)  
  mdat <- getmeth(rgSet)
```

```

beta=getB(mdat)
nmode=nmode(beta, minN = 3,modedist=0.2, nCores = 5)
}

```

---

norm.quantile                      *Quantile normalization.*

---

## Description

Quantile normalization of methylation intensity data across samples for Illumina Infinium Human-Methylation 450 and MethylationEPIC BeadChip.

## Usage

```
norm.quantile(mdat, method = "quantile1")
```

## Arguments

mdat                      An object of class methDataSet or MethylSet.  
method                    Quantile normalization method: "quantile1", "quantile2", or "quantile3".

## Details

By default, method = "quantile1", which will separately quantile normalize Methylated or Unmethylated intensities for Infinium I or II probes. The "quantile2" will quantile normalize combined Methylated or Unmethylated intensities for Infinium I or II probes. The "quantile3" will quantile normalize combined Methylated or Unmethylated intensities for Infinium I and II probes together.

## Value

The output is an an object of class methDataSet or MethylSet.

## Author(s)

Zongli Xu

## References

Pidsley, R., CC, Y.W., Volta, M., Lunnon, K., Mill, J. and Schalkwyk, L.C. (2013) A data-driven approach to preprocessing Illumina 450K methylation array data. BMC genomics, 14, 293.

## Examples

```

#for methDataSet
path <- file.path(find.package("minfiData"), "extdata")
rgSet <- readidat(path = path, recursive = TRUE)
mdat<-preprocessENmix(rgSet, bgParaEst="oob", nCores=6)
mdatq<-norm.quantile(mdat, method="quantile1")

#for MethylSet
if (require(minfiData)) {
mdat=preprocessENmix(RGsetEx, bgParaEst="oob", nCores=6)
}

```

```
mdatq=norm.quantile(mdat,method="quantile1")
}
```

---

 oxBS.MLE

*oxBS-MLE.*


---

### Description

Maximum Likelihood Estimate (MLE) of 5-methylcytosine (5mC) and 5-hydroxymethylcytosine (5hmC) using sequencing/array data from paired bisulfite and oxidative bisulfite treated DNA experiments.

### Usage

```
oxBS.MLE(beta.BS,beta.oxBS,N.BS,N.oxBS)
```

### Arguments

beta.BS	A matrix of methylation beta values (proportion of methylated sites estimated as methylated intensity over total intensity) obtained from bisulfite (BS) experiments
beta.oxBS	A matrix of methylation beta values obtained from oxidative bisulfite (oxBS) experiments
N.BS	A matrix of total signals (sum of methylated and unmethylated intensity values) from BS experiments
N.oxBS	A matrix of total signals from oxBS experiments

### Details

For all the inputs (beta.BS, beta.oxBS, N.BS and N.oxBS), the rows should be corresponding to CpG loci and the columns should be corresponding to samples. The row/column names in all four matrices should be the same. For a specific CpG of a sample, if any one of the four values (beta.BS, beta.oxBS, N.BS and N.oxBS) is NA, or N.BS is zero, or N.oxBS is zero, the MLE of both 5mC and 5hmC levels will be set as NA.

### Value

The output is a list with two elements: 5mC: a matrix of estimated 5mC levels. 5hmC: a matrix for estimated 5hmC levels.

### Author(s)

Liang Niu and Zongli Xu

### References

Zongli Xu, Jack A. Taylor, Yuet-Kin Leung, Shuk-Mei Ho and Liang Niu, *oxBS-MLE: an efficient method to estimate 5-methylcytosine and 5-hydroxymethylcytosine in paired bisulfite and oxidative bisulfite treated DNA*, Bioinformatics. 2016

**Examples**

```
# load example data
load(system.file("oxBS.MLE.RData", package="ENmix"))
# run oxBS.MLE
resu<-oxBS.MLE(beta.BS,beta.oxBS,N.BS,N.oxBS)
dim(resu[["5mC"]])
dim(resu[["5hmC"]])
```

p.qqplot

*P value Q-Q plot***Description**

P value Q-Q plot with optional confidence interval

**Usage**

```
p.qqplot(pvalues, outf="qq", outfmt="jpg", draw.conf=TRUE,
conf.col="lightgray", conf.alpha=.95, pch=20, col="black", reducesize=0, ...)
```

**Arguments**

pvalues	An numeric vector of P values
outf	figure file name, default "qq"
outfmt	Output figure file format, "jpg" or "eps"
draw.conf	Whether to draw confidence interval of expected P values under NULL hypothesis
conf.col	Color code of confidence interval
conf.alpha	Confidence interval range, 0.95 in default
pch	Point type code
col	Point color code
reducesize	A positive interger, larger the value, smaller the eps file size. Smaller file size is achieved by skipping some densely packed data points
...	Arguments pass to plot

**Details**

P value Q-Q plot with optional confidence interval

**Author(s)**

Zongli Xu

**Examples**

```
dat=simubed()
p.qqplot(pvalues=dat$p, draw.conf=TRUE, outf="qq_try", outfmt="jpg")
```

---

pcrplot

*Principal component regression plot*

---

### Description

First, principal component analysis will be performed in the standardized input data matrix (standardized for each row/CpG), and then the specified number of top principal components (which explain most data variation) will be used to perform linear regression with each specified variable. Regression P values will be plotted for exploration of methylation data variance structure or identification of possible confounding variables in association analysis.

### Usage

```
pcrplot(beta, cov, npc=50)
```

### Arguments

beta	A methylation beta value matrix with rows for probes and columns for samples. The input matrix should not contain any missing value.
cov	A data frame of covariates. Categorical variables should be converted to factors. The number of rows should equal to the number of columns in beta matrix
npc	The number of top ranked principal components to be plotted

### Value

A jpeg figure "svdscreepplot.jpg" to show the variations explained by each principal component.

A jpeg figure "pcr\_diag.jpg" to show association strength between principal components and covariates with cell colors indicating different levels of association P values.

### Author(s)

Zongli Xu

### References

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015

### Examples

```
if (require(minfiData)) {  
  mdat <- preprocessRaw(RGsetEx)  
  beta=getBeta(mdat, "Illumina")  
  group=pData(mdat)$Sample_Group  
  slide=factor(pData(mdat)$Slide)  
  cov=data.frame(group,slide)  
  pcrplot(beta,cov,npc=6)  
}
```

---

plotCtrl                      *Internal control plot*

---

### Description

The function will generate a series of internal control plots that are similar to the plots from Illumina GenomeStudio software. Users should refer to GenomeStudio online guide to interpret these figures. These figures can be used to check data quality and experimental procedures.

### Usage

```
plotCtrl(rgSet, IDorder=NULL)
```

### Arguments

rgSet	An object of class rgDataSet or RGChannelSet.
IDorder	A list of sample ids in the order specified by user. The list can be a subset of sample ids in input dataset. If an id list is provided, all plots will be produced in the order of the list.

### Value

A set of internal control figures in jpeg format.

### Author(s)

Zongli Xu

### References

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015.

### Examples

```
if (require(minfiData)) {  
  #rgDataSet as input  
  path <- file.path(find.package("minfiData"), "extdata")  
  rgSet <- readidat(path = path, recursive = TRUE)  
  plotCtrl(rgSet)  
  
  #RGChannelSet as input  
  pinfo=pData(RGsetEx)  
  IDorder=rownames(pinfo)[order(pinfo$Slide, pinfo$Array)]  
  plotCtrl(RGsetEx, IDorder)  
}
```

---

predSex *Estimating sample sex*

---

### Description

Estimating sample sex based on methylation data

### Usage

```
predSex(mdat, cutoff = 2)
```

### Arguments

mdat                    An object of class MethDataSet or rgDataSet.  
 cutoff                 The difference in log<sub>2</sub> total intensity between X and Y chromosomes

### Details

Estimation of sex is based on the difference of log<sub>2</sub> median total intensity measures on the X and Y chromosomes.

### Author(s)

Zongli Xu

### Examples

```
if (require(minfiData)) {
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  sex=predSex(rgSet)
}
```

---

preprocessENmix *The ENmix background correction*

---

### Description

The ENmix background correction for HumanMethylation 450 and MethylationEPIC BeadChip. ENmix models methylation signal intensities with a flexible exponential-normal mixture distribution, and models background noise with a truncated normal distribution. ENmix will split BeadChip intensity data into 6 parts and separately model methylated and unmethylated intensities, 2 different color channels and 2 different probe types.

### Usage

```
preprocessENmix(rgSet, bgParaEst = "oob", dyeCorr="RELIC", QCinfo=NULL, exQCsample=TRUE,
  exQCcpg=TRUE, exSample=NULL, exCpG=NULL, nCores = 2)
```

**Arguments**

rgSet	An object of class rgDataSet, methDataSet, RGChannelSetExtended, RGChannelSet or MethylSet.
bgParaEst	Method to estimate background normal distribution parameters. Options are: "oob", "est", or "neg".
dyeCorr	Dye bias correction method, "mean": correction based on averaged red/green ratio, or "RELIC": correction with RELIC method (default method), or "none": no dye bias correction.
QCinfo	If QCinfo object from function QCinfo() is provided, low quality samples (if exQCsample=TRUE) and CpGs (if exQCcpg=TRUE) will be excluded before background correction.
exQCsample	If TRUE, low quality samples listed in QCinfo will be excluded.
exQCcpg	If TRUE, low quality CpGs listed in QCinfo will be excluded.
exSample	User specified samples to be excluded before background correction
exCpG	User specified probes to be excluded before background correction
nCores	Number of cores will be used for computation

**Details**

By default, ENmix will use out-of-band Infinium I intensities ("oob") to estimate normal distribution parameters for modeling background noise. Option "est" will use combined methylated and unmethylated intensities to estimate background distribution parameters separately for each color channel and each probe type. Option "neg" will use 600 chip internal controls probes to estimate background distribution parameters. If rgSet is a MethylSet, then only option "est" can be selected.

**Value**

An object of class same with input data

**Author(s)**

Zongli Xu and Liang Niu

**References**

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip. *Nucleic Acids Research* 2015.

Zongli Xu, Sabine A. S. Langie, Patrick De Boever, Jack A. Taylor1 and Liang Niu, RELIC: a novel dye-bias correction method for Illumina Methylation BeadChip, *BMC Genomics*. 2017

**Examples**

```
if (require(minfiData)) {
  #rgDataSet as input
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  #quality control information
  qc<-QCinfo(rgSet)
  #further excluding samples which are not in the qc$badsample list
  ex_id=c("5723646053_R04C02")
}
```

```
#further excluding cpGs which are not in the qc$badCpG list
ex_cg=c("cg00000622", "cg00001245", "cg00001261")
mdat=preprocessENmix(rgSet, QCinfo=qc, exSample=ex_id, exCpG=ex_cg, nCores=6)

#RGChannelSet as input
mdat=preprocessENmix(RGsetEx, nCores=6)
}
```

---

 qcfilter
 

---



---

*Filtering out low quality and outlier data*


---

### Description

Outlier was defined as values smaller than 3 times IQR from the lower quartile or greater than 3 times IQR from the upper quartile. If data quality information were provided, low quality data points will be set as missing data first before looking for outliers. All outliers and low quality data will be set as miss in output matrix. If set `impute=TRUE`, imputation will be performed using k-nearest neighbors method to impute all missing values.

### Usage

```
qcfilter(mat, qcscore=NULL, rmoutlier=TRUE, byrow=TRUE, detPthre=0.000001, nbthre=3,
         rmcrc=FALSE, rthre=0.05, cthre=0.05, impute=FALSE, imputebyrow=TRUE, fastimpute=FALSE, ...)
```

### Arguments

<code>mat</code>	An numeric matrix containing methylation beta values
<code>qcscore</code>	If the data quality information (the output from function <code>QCinfo</code> ) were provided, low quality data points as defined by detection p value threshold ( <code>detPthre</code> ) and number of bead threshold ( <code>nbthre</code> ) will be set as missing value.
<code>rmoutlier</code>	if <code>TRUE</code> , outliers data points will be set as missing data <code>NA</code> .
<code>byrow</code>	<code>TRUE</code> : Looking for outliers row by row, or <code>FALSE</code> : column by column.
<code>detPthre</code>	Detection P value threshold to define low quality data points, <code>detPthre=0.000001</code> in default.
<code>nbthre</code>	Number of beads threshold to define low quality data points, <code>nbthre=3</code> in default.
<code>rmcra</code>	<code>TRUE</code> : exclude rows and columns with too many missing values as defined by <code>rthre</code> and <code>cthre</code> . <code>FALSE</code> is in default
<code>rthre</code>	Minimum of percentage of missing values for a row to be excluded
<code>cthre</code>	Minimum of percentage of missing values for a column to be excluded
<code>impute</code>	If <code>TRUE</code> , k-nearest neighbors methods will used for imputation.
<code>imputebyrow</code>	<code>TRUE</code> : impute missing values using similar values in row, or <code>FALSE</code> : in column
<code>fastimpute</code>	If <code>TRUE</code> , probe median will be used for fast imputation.
<code>...</code>	Arguments to be passed to the function <code>impute.knn</code> in R package "impute"

### Value

The output is an numeric matrix.

**Author(s)**

Zongli Xu

**References**

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015.

**Examples**

```

if (require(minfiData)) {
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  qc=QCinfo(rgSet)
  mdat=preprocessENmix(rgSet, QCinfo=qc, nCores=6)
  mdat=norm.quantile(mdat, method="quantile1")
  beta=rcp(mdat)
  #filter out outliers data points only
  b1=qcfilter(beta)
  #filter out low quality and outlier data points
  b2=qcfilter(beta, qcscore=qc)
  #filter out low quality and outlier values, remove rows and columns with
  # too many missing values
  b3=qcfilter(beta, qcscore=qc, rmcr=TRUE)
  #filter out low quality and outlier values, remove rows and columns with
  # too many missing values, and then do imputation
  b3=qcfilter(beta, qcscore=qc, rmcr=TRUE, impute=TRUE)
}

```

QCinfo

*Extract QC information***Description**

Extract information for data quality control: detection P values, number of beads and averaged bisulfite conversion intensity. The function can also identify low quality samples and probes, as well as outlier samples in total intensity or beta value distribution.

**Usage**

```
QCinfo(rgSet, detPthre=0.000001, detPtype="negative", nbthre=3, samplethre=0.05,
       CpGthre=0.05, bisulthre=NULL, outlier=TRUE, distplot=TRUE)
```

**Arguments**

rgSet	An object of class rgDataSet, or RGChannelSetExtended
detPthre	Detection P value threshold to identify low quality data point
detPtype	Calculate detection P values based on negative internal control ("negative") probes or out of the band ("oob") probes
nbthre	Number of bead threshold to identify data point of low quality

samplethre	Threshold to identify samples with low data quality, the percentage of low quality methylation data points across probes for each sample
CpGthre	Threshold to identify probes with low data quality, percentage of low quality methylation data points across samples for each probe
bisulthre	Threshold of bisulfite intensity for identification of low quality samples. By default, Mean - 3 x SD of sample bisulfite control intensities will be used as a threshold.
outlier	If TRUE, outlier samples in total intensity or beta value distribution will be identified and classified as bad samples.
distplot	TRUE or FALSE, whether to produce beta value distribution plots before and after QC.

### Value

detP: a matrix of detection P values

nbead: a matrix for number of beads

bisul: a vector of averaged intensities for bisulfite conversion controls per sample

badsample: a list of low quality or outlier samples

badCpG: a list of low quality CpGs

outlier\_sample: a list of outlier samples in methylation beta value or total intensity distribution.

Figure "qc\_sample.jpg": scatter plot of Percent of low quality data per sample vs. Average bisulfite conversion intensity

Figure "qc\_CpG.jpg": histogram for Percent of low quality data per CpG.

Figure "freqpolygon\_beta\_beforeQC.jpg": distribution plot of input data, samples colored in red are "bad" samples, list in badsample, including samples with low data quality and outlier in methylation beta value or total intensity.

Figure "freqpolygon\_beta\_afterQC.jpg": distribution plot input data after filtering "bad" samples.

### Author(s)

Zongli Xu

### References

Zongli Xu, Liang Niu, Leping Li and Jack A. Taylor, *ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip*. Nucleic Acids Research 2015.

### Examples

```
if (require(minfiData)) {
  #rgDataSet as input
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  qc=QCinfo(rgSet)

  #RGChannelSetExtended as input
  sheet <- read.metharray.sheet(file.path(find.package("minfiData"), "extdata"),
    pattern = "csv$")
  rgSet <- read.metharray.exp(targets = sheet, extended = TRUE)
```

```
qc<-QCinfo(rgSet)
}
```

rcp

*Illumina methylation array probe type bias correction***Description**

Probe design type bias correction using Regression on Correlated Probes (RCP) method

**Usage**

```
rcp(mdat, dist=25, quantile.grid=seq(0.001,0.999,by=0.001), qcscore = NULL,
nbthre=3, detPthre=0.000001)
```

**Arguments**

mdat	An object of class methDataSet or MethylSet.
dist	Maximum distance in base pair between type I and type II probe pairs for regression calibration
quantile.grid	Quantile grid used in linear regression
qcscore	Data quality information object, the output from function QCinfo. If the object is provided, low quality data points as defined by detection p value threshold (detPthre) or number of bead threshold (nbthre) will be set as missing values.
detPthre	Detection P value threshold to define low quality data points
nbthre	Number of beads threshold to define low quality data points, nbthre=3 in default.

**Details**

The function will first identify type I and type II probe pairs within a specified distance, and then perform linear regression calibration between the probe types. With the estimates the function will then adjust type II data using type I data as references.

**Value**

A beta value matrix

**Author(s)**

Liang Niu, Zongli Xu

**References**

Liang Niu, Zongli Xu and Jack A. Taylor *RCP: a novel probe design bias correction method for Illumina Methylation BeadChip, Bioinformatics 2016*

## Examples

```

if (require(minfiData)) {
  #methDataSet as input
  path <- file.path(find.package("minfiData"), "extdata")
  rgSet <- readidat(path = path, recursive = TRUE)
  qc=QCinfo(rgSet)
  mdat=preprocessENmix(rgSet, QCinfo=qc, nCores=6)
  mdat=norm.quantile(mdat, method="quantile1")
  beta=rcp(mdat)

  #methylset as input
  sheet <- read.metharray.sheet(file.path(find.package("minfiData"), "extdata"),
    pattern = "csv$")
  rgSet <- read.metharray.exp(targets = sheet, extended = TRUE)
  qc=QCinfo(rgSet)
  mdat=preprocessENmix(rgSet, QCinfo=qc, nCores=6)
  mdat=norm.quantile(mdat, method="quantile1")
  beta=rcp(mdat)
}

```

---

rcp2

*Modified RCP method*


---

## Description

Modified RCP method to normalize user's data to a list of reference values

## Usage

```
rcp2(datMeth, reference, quantile.grid=seq(0.001, 0.999, by=0.001))
```

## Arguments

datMeth	A matrix with row for probes and column for samples
reference	A data frame with two columns, "cg" for CpG names and "meth_mean" for reference values
quantile.grid	Quantile grid used in linear regression

## Details

The function will normalize user data to a reference value distribution based on common set of probe between user data and reference data.

## Value

A matrix with same dimension of user data

## Author(s)

Zongli Xu

## References

Liang Niu, Zongli Xu and Jack A. Taylor *RCP: a novel probe design bias correction method for Illumina Methylation BeadChip, Bioinformatics 2016*

## Examples

```
require(minfiData)
path <- file.path(find.package("minfiData"), "extdata")
#based on rgDataset
rgSet <- readidat(path = path, recursive = TRUE)
meth=getmeth(rgSet)
beta=getB(meth)
reference=data.frame(cg=rownames(beta), goldstandard=beta[, 3])
reference=reference[sample(1:nrow(reference), 2000), ]
beta2=rcp2(beta, reference)
```

---

readidat

*Parsing IDAT files for Illumina methylation arrays .*

---

## Description

Read in IDAT files and create a rgDataSet with probe annotation

## Usage

```
readidat(path = NULL, manifestfile=NULL, recursive = TRUE, verbose = FALSE, force=FALSE)
```

## Arguments

path	Directory where idat files are located
manifestfile	Array manifestfile, which can be downloaded from Illumina website. Bioconductor manifest package will be used if not provided
recursive	if TRUE, idat files in the subdirectories will also be read in
verbose	if TRUE, detailed running info will be printed on screen
force	if TRUE, arrays with different sizes will be merged together

## Details

Some array types and corresponding manifestfiles can be guessed by the program based on the number of probes per array. However, we recommend to provide correct manifest file directly, which can be easily downloaded from Illumina website, see below for some examples.

Probe annotation info can be extracted using command rowData

## Value

An object of class rgDataSet,

## Author(s)

Zongli Xu

**Examples**

```

#Illumina methylation array manifestfile

#Infinium Mouse Methylation Manifest File (CSV)
system("wget https://support.illumina.com/content/dam/illumina-support/documents/
downloads/productfiles/mouse-methylation/infinium-mouse-methylation-
manifest-file-csv.zip")
system("unzip infinium-mouse-methylation-manifest-file-csv.zip")
mf="infinium-mouse-methylation-manifest-file.csv"

#for MethylationEPIC v2.0
#download InfiniumMethylationEPICv2.0ProductFiles
#mf= path to file "EPIC-8v2-0_A1.csv" or file "EPIC-8v2-0_A2.csv"
#manifest=readmanifest(mf)

#for MethylationEPIC v1.0 B5
system("wget https://webdata.illumina.com/downloads/productfiles/
methylationEPIC/infinium-methylationepic-v-1-0-b5-
manifest-file-csv.zip")
system("unzip infinium-methylationepic-v-1-0-b5-manifest-file-csv.zip")
mf="infinium-methylationepic-v-1-0-b5-manifest-file.csv"
manifest=readmanifest(mf)

#for MethylationEPIC v1.0 B4
system("wget https://webdata.illumina.com/downloads/productfiles/
methylationEPIC/infinium-methylationepic-v-1-0-b4-
manifest-file-csv.zip")
system("unzip infinium-methylationepic-v-1-0-b4-manifest-file-csv.zip")
mf="MethylationEPIC_v-1-0_B4.csv"
manifest=readmanifest(mf)

#for HumanMethylation450
system("wget https://webdata.illumina.com/downloads/productfiles/
humanmethylation450/humanmethylation450_15017482_v1-2.csv")
mf="HumanMethylation450_15017482_v1-2.csv"

mf="HumanMethylation450_15017482_v1-2.csv"
if(require(minfiData)){
path <- file.path(find.package("minfiData"),"extdata")
rgSet <- readidat(path = path,manifestfile=mf,recursive = TRUE)

#without providing manifest file, corresponding R manifestfile package will be used
rgSet <- readidat(path = path,recursive = TRUE)
}

```

---

readmanifest

*Parsing Illumina methylation arrays manifest file.*


---

**Description**

Parsing Illumina methylation arrays manifest file.

**Usage**

```
readmanifest(file)
```

**Arguments**

file                    Illumina methylation array manifest file, downloaded from Illuminal website

**Value**

An object of dataframe caintaining probe annotation information

**Author(s)**

Zongli Xu

**Examples**

```
#for MethylationEPIC v1.0 B5
system("wget https://webdata.illumina.com/downloads/productfiles/methylationEPIC/
       infinium-methylationepic-v-1-0-b5-manifest-file-csv.zip")
system("unzip infinium-methylationepic-v-1-0-b5-manifest-file-csv.zip")
mf="infinium-methylationepic-v-1-0-b5-manifest-file.csv"
manifest=readmanifest(mf)

#for MethylationEPIC v1.0 B4
system("wget https://webdata.illumina.com/downloads/productfiles/methylationEPIC/
       infinium-methylationepic-v-1-0-b4-manifest-file-csv.zip")
system("unzip infinium-methylationepic-v-1-0-b4-manifest-file-csv.zip")
mf="MethylationEPIC_v-1-0_B4.csv"
manifest=readmanifest(mf)

#for HumanMethylation450
system("wget https://webdata.illumina.com/downloads/productfiles/
       humanmethylation450/humanmethylation450_15017482_v1-2.csv")
mf="HumanMethylation450_15017482_v1-2.csv"
manifest=readmanifest(mf)
```

---

relic

*REgression on Logarithm of Internal Control probes (RELIC)*

---

**Description**

REgression on Logarithm of Internal Control probes (RELIC) correct for dye bias on whole array by utilizing the intensity values of paired internal control probes that monitor the two color channels.

**Usage**

```
relic (mdat, at_red, cg_grn)
```

**Arguments**

<code>mdat</code>	An object of class <code>methDataSet</code> or <code>MethylSet</code> .
<code>at_red</code>	an intensity matrix for Illumina control probes "NORM_A" and "NORM_T"
<code>cg_grn</code>	an intensity matrix for Illumina control probes "NORM_C" and "NORM_G"

**Details**

The Illumina MethylationEPIC BeadChip contains 85 pairs of internal normalization control probes (name with prefix NORM\_A, NORM\_T, NORM\_G or NORM\_C), while its predecessor, Illumina HumanMethyl-ation450 BeadChip contains 93 pairs. RELIC first performs a regression on the logarithms of the intensity values of the normalization control probes to derive a quantitative relationship between red and green channels, and then uses the relationship to correct for dye-bias on intensity values for whole array.

**Value**

An object of class `methDataSet` or `MethylSet` depends on input class.

**Author(s)**

Zongli Xu and Liang Niu

**References**

Zongli Xu, Sabine A. S. Langie, Patrick De Boever, Jack A. Taylor and Liang Niu, RELIC: a novel dye-bias correction method for Illumina Methylation BeadChip, *BMC Genomics*. 2017

**See Also**

Package `preprocessENmix`

**Examples**

```
if (require(minfiData)) {

##background correction and dye bias correction
#rgDataSet as input
path <- file.path(find.package("minfiData"), "extdata")
rgSet <- readidat(path = path, recursive = TRUE)
mdat <- preprocessENmix(rgSet, bgParaEst="oob", nCores=6, dyeCorr = "RELIC")

#RGChannelSet as input
mdat=preprocessENmix(RGsetEx, bgParaEst="oob", nCores=6, dyeCorr = "RELIC")

##dye bias correction only
#methDataSet as input
path <- file.path(find.package("minfiData"), "extdata")
rgSet <- readidat(path = path, recursive = TRUE)
ctrls <- getCGinfo(rgSet, type="ctrl")
ctrls <- ctrl[ctrls$Address %in% rownames(rgSet),]

ctrl_r <- assays(rgSet)$Red[ctrls$Address,]
ctrl_g <- assays(rgSet)$Green[ctrls$Address,]
CG.controls <- ctrl$Type %in% c("NORM_C", "NORM_G")
```

```

AT.controls <- ctrls$Type %in% c("NORM_A", "NORM_T")
cg_grn=ctrl_g[CG.controls,]
at_red=ctrl_r[AT.controls,]
rownames(cg_grn) = ctrls$ExtendedType[CG.controls]
rownames(at_red) = ctrls$ExtendedType[AT.controls]
mdat=getmeth(rgSet)
mdat <- relic(mdat,at_red,cg_grn)

#MethylSet as input
ctrls <- getProbeInfo(RGsetEx,type="Control")
ctrls <- ctrls[ctrls$Address %in% featureNames(RGsetEx),]
ctrl_r <- getRed(RGsetEx)[ctrls$Address,]
ctrl_g <- getGreen(RGsetEx)[ctrls$Address,]
CG.controls <- ctrls$Type %in% c("NORM_C","NORM_G")
AT.controls <- ctrls$Type %in% c("NORM_A","NORM_T")
cg_grn <- ctrl_g[CG.controls,]
at_red <- ctrl_r[AT.controls,]
rownames(cg_grn) = ctrls$ExtendedType[CG.controls]
rownames(at_red) = ctrls$ExtendedType[AT.controls]
mdat <- preprocessRaw(RGsetEx)
mdat <- relic(mdat,at_red,cg_grn)

}

```

---

repicc

*Calculating intraclass correlation coefficient using replicate samples*


---

## Description

The function can be used to calculate ICC for each CpG probe using balanced or unbalanced replicate samples.

## Usage

```

repicc(dat,repid,mvalue=FALSE,nCores=2,qcflag=FALSE,qc=NULL,
       detPthre=0.05,nbthre=3)

```

## Arguments

dat	Methylation beta value matrix
repid	A data frame with two variables, id and idx. id should be the same with column name of "dat", idx is a variable to show the relationship between samples with same value for samples from same individual.
mvalue	If TRUE, the beta value will be converted to M value for calculation of ICC
nCores	Number of cores will be used for calculation of ICC
qcflag	Whether to perform QC before calculation of ICC
qc	QC object from function QCinfo
detPthre	If qcflag=TRUE, the methylation values with detection P value higher than the threshold will be removed before calculation
nbthre	If qcflag=TRUE, the methylation values with number of bead smaller

**Value**

A data frame containing ICC for each probe

**Author(s)**

Zongli Xu

**References**

Zongli Xu, Jack A Taylor. *Reliability of DNA methylation measures using Illumina methylation BeadChip*. Epigenetics 2020

**Examples**

```
if (require(minfiData)){
  path <- file.path(find.package("minfiData"),"extdata")
  rgSet <- readidat(path = path,recursive = TRUE)
  mdat=getmeth(rgSet)
  beta=getB(mdat,"Illumina")
  repid=data.frame(id=c("5723646052_R02C02", "5723646052_R04C01", "5723646052_R05C02",
    "5723646053_R04C02", "5723646053_R05C02", "5723646053_R06C02"),idx=c(1,1,2,2,2,2))
  iccresu<-repicc(dat=beta,repid=repid)
}
```

---

 rgDataSet-class

 Class "rgDataSet"

---

**Description**

A class for storing Illumina methylation array raw intensity data of two color channels, and probe annotation information.

**Usage**

```
rgDataSet(Red = new("matrix"), Green = new("matrix"),
  NBeads = new("matrix"),rowData=new("DataFrame"),ictrl= new("DataFrame"),...)
```

**Arguments**

Red	A matrix of Red channel intensity values with row for methylation probes and column for samples
Green	A matrix of Green channel intensity values with row for methylation probes and column for samples
NBeads	A matrix contains the number of beads used to generate intensity values on the Red and Green channels.
rowData	A dataframe contains probe annotation information
ictrl	A dataframe contains detailed information for Illumina internal control probes
...	other arguments for class SummarizedExperiment

**Value**

An object of class rgDataSet

**Examples**

```
showClass("rgDataSet")
```

---

rm.cgsuffix	<i>Remove suffix from CpG names and combine duplicated CpGs</i>
-------------	---

---

**Description**

Remove suffix from CpG names for EPIC v2 BeadChips and combined values for duplicated CpGs

**Usage**

```
rm.cgsuffix(datMeth)
```

**Arguments**

datMeth      A methlation data matrix with row names for CpG id

**Details**

Remove suffix from CpG names for EPIC v2 BeadChips and combined values for duplicated CpGs

**Value**

A matrix with number of rows equal or less than input data.

**Author(s)**

Zongli Xu

**Examples**

```
#beta matrix with row for CpGs and column for samples  
beta2=rm.cgsuffix(beta)
```

---

simubed

*Simulation of bed format example file.*

---

**Description**

Simulation of bed format example file.

**Usage**

```
simubed(nprobe=1000)
```

**Arguments**

nprobe            Number of probes on each chromosome, default is 1000

**Details**

Simulation of bed format example file.

**Value**

A data frame

**Author(s)**

Zongli Xu

**Examples**

```
simubed(nprobe=1000)
```

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