

Package ‘HERON’

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Description HERON is a software package for analyzing peptide binding array data. In addition to identifying significant binding probes, HERON also provides functions for finding epitopes (string of consecutive peptides within a protein). HERON also calculates significance on the probe, epitope, and protein level by employing meta p-value methods. HERON is designed for obtaining calls on the sample level and calculates fractions of hits for different conditions.

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BugReports <https://github.com/Ong-Research/HERON/issues>

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Author Sean McIlwain [aut, cre] (ORCID:

<<https://orcid.org/0000-0002-3820-8400>>),

Irene Ong [aut] (ORCID: <<https://orcid.org/0000-0002-9353-6941>>)

Maintainer Sean McIlwain <sean.mcilwain@wisc.edu>

Contents

HERON-package	3
addSequenceAnnotations	3
calcCombPValues	4
calcEpitopePValues	5
calcProbePValuesTPaired	6
calcProbePValuesTUnpaired	7
calcProbePValuesWUnpaired	7
calcProteinPValues	8
catSequences	9
convertSequenceDSToProbeDS	9
findBlocksProbeT	10
findBlocksT	10
findEpitopeSegments	11
getEpitopeID	12
getEpitopeIDsToProbeIDs	12
getEpitopeProbeIDs	13
getEpitopeProtein	13
getEpitopeStart	14
getEpitopeStop	14
getKofN	15
getProteinLabel	15
getProteinStart	16
getProteinTiling	16
heffron2021_wuhan	17
HERONEpitopeDataSet-class	17
HERONProbeDataSet-class	18
HERONProteinDataSet-class	18
HERONSequenceDataSet-class	19
log2Transform	20
makeEpitopeCalls	20
makeProbeCalls	21
makeProteinCalls	22
min_max	22
oneHitEpitopes	23
oneHitProbes	24
oneProbeEpitopes	24
probeHitSupported	25
pvalue_to_zscore	25
quantileNormalize	26
smoothProbeDS	26

Index

27

HERON-package

HERON: Hierarchical Epitope pROtein biNding

Description

HERON is a software package for analyzing peptide binding array data. In addition to identifying significant binding probes, HERON also provides functions for finding epitopes (string of consecutive peptides within a protein). HERON also calculates significance on the probe, epitope, and protein level by employing meta p-value methods. HERON is designed for obtaining calls on the sample level and calculates fractions of hits for different conditions.

Author(s)

Maintainer: Sean McIlwain <sean.mcilwain@wisc.edu> ([ORCID](#))

Authors:

- Irene Ong <irene.ong@wisc.edu> ([ORCID](#))

See Also

Useful links:

- <https://github.com/Ong-Research/HERON>
- Report bugs at <https://github.com/Ong-Research/HERON/issues>

addSequenceAnnotations

Add Sequence Annotations for Epitopes

Description

Add Sequence Annotations for Epitopes

Usage

```
addSequenceAnnotations(eds)
```

Arguments

eds HERONEpitopeDataSet with probe_meta in metadata()

Value

HERONEpitopeDataSet with the rowData() set with sequence annotations

Examples

```

data(heffron2021_wuhan)
pval_seq_res <- calcCombPValues(heffron2021_wuhan)
pval_pr_res <- convertSequenceDSToProbeDS(pval_seq_res)
calls_res <- makeProbeCalls(pval_pr_res)
segments_res <- findEpitopeSegments(calls_res, "unique")
epval_res <- calcEpitopePValues(calls_res, segments_res)
epval_res <- addSequenceAnnotations(epval_res)

```

calcCombPValues	<i>Calculate p-values using the "exprs" assay</i>
-----------------	---

Description

Calculate p-values using the "exprs" assay

Usage

```

calcCombPValues(
  obj,
  colData_in = NULL,
  d_sd_shift = NA,
  d_abs_shift = NA,
  d_paired = FALSE,
  g_sd_shift = 0,
  use = "tz",
  p_adjust_method = "BH"
)

```

Arguments

obj	HERONSequenceDataSet or HERONProbeDataSet
colData_in	optional column DataFrame (default: NULL => colData(obj))
d_sd_shift	standard deviation shift for differential test
d_abs_shift	absolute shift for differential test
d_paired	run paired analysis
g_sd_shift	standard deviation shift for global test
use	use global-test ("z"), differential-test using t.test ("t"), differential-test using wilcox ("w"), or both global and differential ("tz")
p_adjust_method	method for adjusting p-values

Value

HERONSequenceDataSet/HERONProbeDataSet with the pvalue assay added

Examples

```

data(heffron2021_wuhan)
seq_pval_res <- calcCombPValues(heffron2021_wuhan)

```

calcEpitopePValues *Calculate epitope-level p-values*

Description

Calculate epitope-level p-values

Usage

```
calcEpitopePValues(  
  probe_pds,  
  epitope_ids,  
  metap_method = "wmax1",  
  p_adjust_method = "BH"  
)
```

Arguments

probe_pds HERONProbeDataSet with the "pvalue" assay
epitope_ids vector of epitope ids
metap_method meta p-value method to use (see below)
p_adjust_method what p.adjust method to use.

Details

The meta p-value methods supported by calcEpitopePValues are: min_bonf*, min*, max*, fisher/sumlog, hmp/harmonicmeanp, wilkinsons_min1/tippets, wilkinsons_min2/wmin2, wilkinsons_min3, wilkinsons_min4, wilkinsons_min5, wilkinsons_max1/wmax1, wilkinsons_max2/wmax2, and cct.

When choosing a p-value method, keep in mind that the epitope p-value should be one that requires most of the probe p-values to be small (e.g. *wmax1*) Other p-value methods such as the*cct* and the *hmp* have been shown to be more accurate with p-value that have dependencies.

Value

HERONEpitopeDataSet with "pvalue" and "padj" assays

See Also

[stats::p.adjust()] for p_adjust_parameter.

Examples

```
data(heffron2021_wuhan)  
pval_seq_res <- calcCombPValues(heffron2021_wuhan)  
pval_pr_res <- convertSequenceDSToProbeDS(pval_seq_res)  
calls_res <- makeProbeCalls(pval_pr_res)  
segments_res <- findEpitopeSegments(calls_res, "unique")  
epval_res <- calcEpitopePValues(calls_res, segments_res)
```

`calcProbePValuesTPaired`*Calculate Probe p-values using a differential paired t-test*

Description

Calculate Probe p-values using a differential paired t-test

Usage

```
calcProbePValuesTPaired(  
  probe_mat,  
  colData_in,  
  sd_shift = NA,  
  abs_shift = NA,  
  debug = FALSE  
)
```

Arguments

<code>probe_mat</code>	numeric matrix or data.frame of values
<code>colData_in</code>	design data.frame
<code>sd_shift</code>	standard deviation shift to use when calculating p-values. Either <code>sd_shift</code> or <code>abs_shift</code> should be set
<code>abs_shift</code>	absolute shift to use when calculating p-values.
<code>debug</code>	print debugging information

Value

matrix of p-values on the post columns defined in the `colData` matrix. Attributes of the matrix are:
`pars` - data.frame parameters used in the paired t-test for each row (e.g. `df`, `sd`)
`mapping` - data.frame of mapping used for pre-post column calculation
`diff_mat` - data.frame containing the post-pre differences for each sample (column) and probe (row)

Examples

```
data(heffron2021_wuhan)  
colData_wu <- colData(heffron2021_wuhan)  
pre_idx = which(colData_wu$visit == "pre")  
## Make some samples paired  
colData_post = colData_wu[colData_wu$visit == "post",]  
new_ids = rownames(colData_post)[seq_len(5)]  
colData_wu$ptid[pre_idx[seq_len(5)]] = new_ids  
exprs <- assay(heffron2021_wuhan, "exprs")  
pval_res <- calcProbePValuesTPaired(exprs, colData_wu)
```

`calcProbePValuesTUnpaired`*Calculate Probe p-values using a differential unpaired t-test*

Description

Calculate Probe p-values using a differential unpaired t-test

Usage

```
calcProbePValuesTUnpaired(probe_mat, colData_in, sd_shift = NA, abs_shift = NA)
```

Arguments

<code>probe_mat</code>	numeric matrix or data.frame of values
<code>colData_in</code>	design data.frame
<code>sd_shift</code>	standard deviation shift to use when calculating p-values Either <code>sd_shift</code> or <code>abs_shift</code> should be set
<code>abs_shift</code>	absolute shift to use when calculating p-values

Value

matrix of p-values on the post columns defined in the `colData` matrix

Examples

```
data(heffron2021_wuhan)
colData_wu <- colData(heffron2021_wuhan)
pval_res <- calcProbePValuesTUnpaired(assay(heffron2021_wuhan), colData_wu)
```

`calcProbePValuesWUnpaired`*Calculate Probe p-values using a two-sample wilcoxon test*

Description

Calculate Probe p-values using a two-sample wilcoxon test

Usage

```
calcProbePValuesWUnpaired(probe_mat, colData_in, exact = NULL, abs_shift = 0)
```

Arguments

<code>probe_mat</code>	numeric matrix or data.frame of values
<code>colData_in</code>	design data.frame
<code>exact</code>	a logical indicating whether an exact p-value should be computed (see <code>wilcox.test</code> for details)
<code>abs_shift</code>	absolute shift to use when calculating p-values

Value

matrix of p-values on the post columns defined in the colData matrix

Examples

```
data(heffron2021_wuhan)
colData_wu <- colData(heffron2021_wuhan)
pval_res <- calcProbePValuesWUnpaired(assay(heffron2021_wuhan), colData_wu)
```

calcProteinPValues *Calculate protein-level p-values*

Description

Calculate protein-level p-values

Usage

```
calcProteinPValues(epitope_ds, metap_method = "wmin1", p_adjust_method = "BH")
```

Arguments

epitope_ds	HERONEpitopeDataSet with the "pvalue" assay
metap_method	meta p-value method to use
p_adjust_method	p.adjust method to use

Details

see calcEpitopePValues for a list of meta p-value methods supported by HERON. the protein should be one that requires at least one of the epitope p-values to be small (e.g. wmax1).

Value

HERONProteinDataSet with the "pvalue" and "padj" assays

See Also

[stats::p.adjust()] for p_adjust_parameter.
[calcEpitopePValues()] for meta p-value methods

Examples

```
data(heffron2021_wuhan)
pval_seq_res <- calcCombPValues(heffron2021_wuhan)
pval_pr_res <- convertSequenceDSToProbeDS(pval_seq_res)
calls_res <- makeProbeCalls(pval_pr_res)
segments_res <- findEpitopeSegments(calls_res, "unique")
epval_res <- calcEpitopePValues(calls_res, segments_res)
ppval_res <- calcProteinPValues(epval_res)
```

catSequences	<i>Concatenate sequences together based upon their start positions. Assumes the probe sequences have an overlap.</i>
--------------	--

Description

Concatenate sequences together based upon their start positions. Assumes the probe sequences have an overlap.

Usage

```
catSequences(positions, sequences)
```

Arguments

positions	start positions of probes in protein
sequences	probe sequences of probes

Value

concatenated sequence (character)

Examples

```
positions <- c(1,2)
sequences <- c("MSGASFEFGVFSPLYL", "SGSASFEFGVFSPLYLT")
catSequences(positions, sequences)
```

convertSequenceDSToProbeDS

Convert HERONSequenceDataSet to HERONProbeDataSet

Description

Convert HERONSequenceDataSet to HERONProbeDataSet

Usage

```
convertSequenceDSToProbeDS(seq_ds, probe_meta)
```

Arguments

seq_ds	a HERONSequenceDataSet object
probe_meta	optional data.frame with the PROBE_SEQUENCE, PROBE_ID columns the probe meta data frame can be provided within the metadata()\$probe_meta or as a argument to the function. The argument supersedes the metadata list.

Value

HERONProbeDataSet

Examples

```
data(heffron2021_wuhan)
probe_ds <- convertSequenceDSToProbeDS(heffron2021_wuhan)
probe_meta <- metadata(heffron2021_wuhan)$probe_meta
probe_ds <- convertSequenceDSToProbeDS(heffron2021_wuhan, probe_meta)
```

findBlocksProbeT	<i>Find Blocks of consecutive probes</i>
------------------	--

Description

This function will find blocks of consecutive probes within the passed probe parameter

Usage

```
findBlocksProbeT(
  probes,
  protein_tiling,
  proteins = getProteinLabel(probes),
  starts = getProteinStart(probes)
)
```

Arguments

probes	vector of probe identifiers of the format c(Prot1;1, ... Prot1;10)
protein_tiling	tiling of the associated proteins
proteins	associated proteins to probes (cache speed up)
starts	associated starts from probes (cache speed up)

Value

data.frame with the Protein, Start, Stop, and Number.Of.Probes columns

Examples

```
findBlocksProbeT(c("A;1", "A;2", "A;3", "B;2", "B;3", "C;10", "A;5", "A;6"))
```

findBlocksT	<i>Find consecutive probes</i>
-------------	--------------------------------

Description

Find consecutive probes

Usage

```
findBlocksT(prot_df, protein_tiling)
```

Arguments

prot_df data.frame with the Protein and Starting position of the probe
protein_tiling tiling for information for each protein

Value

data.frame with the Protein, Start, Stop, and Number.Of.Probes columns

Examples

```
probes = c("A;1","A;2","A;3", "A;5","A;6", "A;8")
prot_df = data.frame(
  Protein = getProteinLabel(probes),
  Pos = getProteinStart(probes)
)
findBlocksT(prot_df)
```

findEpitopeSegments *Find Epitopes from probe stats and calls.*

Description

Find Epitopes from probe stats and calls.

Usage

```
findEpitopeSegments(
  PDS_obj,
  segment_method = "unique",
  segment_score_type = "binary",
  segment_dist_method = "hamming",
  segment_cutoff = "silhouette"
)
```

Arguments

PDS_obj HERONProbeDataSet with pvalues and calls in the assay
segment_method which epitope finding method to use (binary or zscore, applies for hclust or skater)
segment_score_type which type of scoring to use for probes
segment_dist_method what kind of distance score method to use
segment_cutoff for clustering methods, what cutoff to use (either numeric value or 'silhouette')

Value

a vector of epitope identifiers or segments found

Examples

```
data(heffron2021_wuhan)
seq_pval_res <- calcCombPValues(heffron2021_wuhan)
pr_pval_res <- convertSequenceDSToProbeDS(seq_pval_res)
pr_calls_res <- makeProbeCalls(pr_pval_res)
segments_res <- findEpitopeSegments(pr_calls_res)
```

getEpitopeID	<i>Create EpitopeID from protein, first and last probes</i>
--------------	---

Description

Create EpitopeID from protein, first and last probes

Usage

```
getEpitopeID(protein, start, stop)
```

Arguments

protein	vector of proteins
start	vector of first probe protein start positions
stop	vector of last probe protein start positions

Value

vector of epitope ids

Examples

```
getEpitopeID("A", 1, 2)
```

getEpitopeIDsToProbeIDs	<i>Get probe ids from a vector of epitope ids</i>
-------------------------	---

Description

Get probe ids from a vector of epitope ids

Usage

```
getEpitopeIDsToProbeIDs(epitope_ids, tiling = 1)
```

Arguments

epitope_ids	vector of epitope identifiers
tiling	tiling of probes across proteins

Value

data.frame of epitope_to_probe mappings

Examples

```
getEpitopeIDsToProbeIDs(c("A_1_5", "C_8_12"))
```

getEpitopeProbeIDs *Get the vector of probes from an epitope id*

Description

Get the vector of probes from an epitope id

Usage

```
getEpitopeProbeIDs(epitope_id, tiling = 1)
```

Arguments

epitope_id EpitopeID to obtain probes from
tiling Tiling of the probes across the protein (default 1)

Value

vector of probe_ids that are contained within the epitope

Examples

```
getEpitopeProbeIDs("A_1_5")
```

getEpitopeProtein *Obtain Protein Id from Epitope ID*

Description

Format of EpitopeID is A_B_C, where A is the protein label B is the protein start position of the first probe in the epitope and C is the protein start position of the last probe in the epitope.

Usage

```
getEpitopeProtein(epitope_ids)
```

Arguments

epitope_ids vector of epitope identifier character strings

Value

vector of protein labels

Examples

```
getEpitopeProtein("Prot1_1_5")
```

getEpitopeStart	<i>Obtain first probe's protein start position from Epitope ID</i>
-----------------	--

Description

Obtain first probe's protein start position from Epitope ID

Usage

```
getEpitopeStart(epitope_ids)
```

Arguments

epitope_ids vector of epitope ids

Value

vector of integers indicating first probe start positions in the epitope(s)

Examples

```
getEpitopeStart("Prot1_1_5")
```

getEpitopeStop	<i>Obtain last probe's protein start position from EpitopeID</i>
----------------	--

Description

Obtain last probe's protein start position from EpitopeID

Usage

```
getEpitopeStop(epitope_ids)
```

Arguments

epitope_ids vector of epitope ids

Value

vector of integers indicating the last probe protein start position

Examples

```
getEpitopeStop("Prot1_1_5")
```

getKofN	<i>Get K of N statistics from an experiment with padj and calls</i>
---------	---

Description

Calculates the number of samples (K), the frequency of samples (F), and the percentage of samples (P) called. If the colData DataFrame contains a condition column with at least two conditions, then a K, F, and P is calculated for each condition and the results are reported as separate columns.

Usage

```
getKofN(obj)
```

Arguments

obj HERON Dataset with a "calls" assay

Value

DataFrame with K (#calls), F (fraction calls), P (

Examples

```
data(heffron2021_wuhan)
seq_pval_res <- calcCombPValues(heffron2021_wuhan)
pr_pval_res <- convertSequenceDSToProbeDS(seq_pval_res)
pr_calls_res <- makeProbeCalls(pr_pval_res)
getKofN(pr_calls_res)
```

getProteinLabel	<i>Get Protein Label from Probe</i>
-----------------	-------------------------------------

Description

Get Protein Label from Probe

Usage

```
getProteinLabel(probes)
```

Arguments

probes vector of probes (i.e. c("A;1", "A;2"))

Value

vector of strings indicating the protein associated with the respective probes

Examples

```
getProteinLabel("A;1")
getProteinLabel("B;2")
getProteinLabel(c("A;1", "B;2"))
```

getProteinStart	<i>Get the amino-acid starting position of the probe within the protein.</i>
-----------------	--

Description

Get the amino-acid starting position of the probe within the protein.

Usage

```
getProteinStart(probes)
```

Arguments

probes vector of probes (i.e. c("A;1", "A;2"))

Value

starting locations of the probes with their associated proteins

Examples

```
getProteinStart("A;1")
getProteinStart("B;2")
getProteinStart(c("A;1", "B;2"))
```

getProteinTiling	<i>Get Protein Tiling</i>
------------------	---------------------------

Description

Given a set of probes, estimate the tiling of the probes across the protein. Usually, you will want to calculate this on all the probes available in the dataset.

Usage

```
getProteinTiling(probes, return.vector = TRUE)
```

Arguments

probes vector of probes (i.e. A;1, A;2)
return.vector Return result as vector or return as data.frame

Value

For each protein, the estimating tiling (spacing) of the probes across the amino acid sequence.

Examples

```
getProteinTiling(c("A;1", "A;2", "A;3", "B;2", "B;3", "C;1", "C;3"))
```

heffron2021_wuhan *SARS CoV-2 Wuhan Peptide Binding Array Data*

Description

A subset of data from the paper <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8245122/> publication.

Usage

```
data(heffron2021_wuhan)
```

Format

'heffron2021_wuhan' A HERONSequenceDataSet with and "exprs" assay DataFrame with 1945 rows and 60 columns. Each column is a pre-processed binding signal from a serum sample peptide array set for the SARS-CoV-2. The matrix is a subset of the full matrix and contains sequences from the membrane, envelope, surface (spike), and nucleocapsid proteins.

The metadata()\$probe_meta is a data frame with 1945 rows and 6 columns. The columns are POSITION - starting position of probe within protein, PROBE_SEQUENCE - amino acid sequence of probe, SEQ_ID - protein identifier SEQ_NAME - name of protein, PROBE_ID - combination of protein identifier and starting position, e.g. prot1;5.

The colData() is a DataFrame with 60 rows and 2 columns. The columns are SampleName - name of the sample, visit - either pre or post, ptid - subject id, and condition - all COVID

Value

HERONSequenceDataSet

Source

<https://github.com/Ong-Research/UW_Adult_Covid-19>

HERONEpitopeDataSet-class

HERONEpitopeDataSet object and constructors

Description

HERONEpitopeDataSet is a subclass of SummarizedExperiment used to hold assay information on the epitope-level

Usage

```
HERONEpitopeDataSet(pvalue, ...)
```

Arguments

pvalue	calculate epitope p-value matrix
...	arguments provided to SummarizedExperiment, including metadata

Value

HERONEpitopeDataSet object

Examples

```
pval <- matrix(runif(100),ncol=4)
HERONEpitopeDataSet(pvalue = pval)
```

HERONProbeDataSet-class

HERONProbeDataSet object and constructors

Description

HERONProbeDataSet is a subclass of RangedSummarizedExperiment used to hold assay information on the probe level

Usage

```
HERONProbeDataSet(...)
```

Arguments

... arguments provided to SummarizedExperiment, including metadata.

Value

HERONProbeDataSet object

Examples

```
pds <- HERONProbeDataSet()
```

HERONProteinDataSet-class

HERONProteinDataSet object and constructors

Description

HERONProteinDataSet is a subclass of SummarizedExperiment used to hold assay information on the protein-level

Usage

```
HERONProteinDataSet(pvalue, ...)
```

Arguments

pvalue calculated protein p-value matrix
 ... arguments provided to SummarizedExperiment, including metadata

Value

HERONProteinDataSet object

Examples

```
pval <- matrix(runif(100), ncol=4)
HERONProteinDataSet(pvalue = pval)
```

HERONSequenceDataSet-class

HERONSequenceDataSet object and constructors

Description

HERONSequenceDataSet is a subclass of SummarizedExperiment, used to store the expression values, intermediate calculations, and results of a differential binding code on the seeuqnce-level.

Usage

```
HERONSequenceDataSet(exprs, ...)
```

Arguments

<code>exprs</code>	binding values with rows as sequences and columns as samples
<code>...</code>	arguments provided to SummarizedExperiment, including metadata metadata can contain a probe DataFrame, that maps sequences (column PROBE_SEQUENCE) to probe identifiers (column PROBE_ID)

Value

HERONSequenceDataSet object

Examples

```
exprs <- matrix(seq_len(100),ncol=4)
colnames(exprs) <- c("C1", "C2", "C3", "C4")
sds <- HERONSequenceDataSet(exprs = exprs)
```

log2Transform	<i>log2 transform the "exprs" assay</i>
---------------	---

Description

log2 transform the "exprs" assay

Usage

```
log2Transform(se)
```

Arguments

se SummarizedExperiment with "exprs" assay

Value

SummarizedExperiment with "exprs" assay log2 transformed

Examples

```
data(heffron2021_wuhan)
assay(heffron2021_wuhan, "exprs") <- 2^assay(heffron2021_wuhan, "exprs")
res <- log2Transform(heffron2021_wuhan)
```

makeEpitopeCalls	<i>Make Epitope Calls</i>
------------------	---------------------------

Description

Make Epitope Calls

Usage

```
makeEpitopeCalls(epi_ds, padj_cutoff = 0.05, one_hit_filter = TRUE)
```

Arguments

epi_ds HERONEpitopeDataSet with pvalue assay

padj_cutoff p-value cutoff to use

one_hit_filter filter one hit epitopes?

Value

HERONEpitopeDataSet with calls assay added

Examples

```
data(heffron2021_wuhan)
seq_pval_res <- calcCombPValues(heffron2021_wuhan)
pr_pval_res <- convertSequenceDSToProbeDS(seq_pval_res)
pr_calls_res <- makeProbeCalls(pr_pval_res)
epi_segments_uniq_res <- findEpitopeSegments(
  PDS_obj = pr_calls_res,
  segment_method = "unique"
)
epi_padj_uniq <- calcEpitopePValues(
  probe_pds = pr_calls_res,
  epitope_ids = epi_segments_uniq_res,
  metap_method = "wilkinsons_max1"
)
makeEpitopeCalls(epi_padj_uniq)
```

makeProbeCalls	<i>Making Probe-level Calls</i>
----------------	---------------------------------

Description

makeProbeCalls returns call information on a HERONProbeDataSet using the "padj" assay

Usage

```
makeProbeCalls(pds, padj_cutoff = 0.05, one_hit_filter = TRUE)
```

Arguments

pds HERONProbeDataSet with the "padj" assay
padj_cutoff cutoff to use
one_hit_filter filter out one-hit probes?

Value

HERONProbeDataSet with the "calls" assay added

Examples

```
data(heffron2021_wuhan)
pval_seq_res <- calcCombPValues(heffron2021_wuhan)
pval_probe_res <- convertSequenceDSToProbeDS(pval_seq_res)
calls_res <- makeProbeCalls(pval_probe_res)
```

makeProteinCalls *Make Protein-level Calls*

Description

Make Protein-level Calls

Usage

```
makeProteinCalls(prot_ds, padj_cutoff = 0.05, one_hit_filter = FALSE)
```

Arguments

prot_ds HERONProteinDataSet with the "padj" assay
 padj_cutoff cutoff to use
 one_hit_filter use the one-hit filter?

Value

HERONProteinDataSet with the "calls" assay added

Examples

```
data(heffron2021_wuhan)
seq_pval_res <- calcCombPValues(heffron2021_wuhan)
pr_pval_res <- convertSequenceDSToProbeDS(seq_pval_res)
pr_calls_res <- makeProbeCalls(pr_pval_res)
epi_segments_uniq_res <- findEpitopeSegments(
  PDS_obj = pr_calls_res,
  segment_method = "unique"
)
epi_padj_uniq <- calcEpitopePValues(
  probe_pds = pr_calls_res,
  epitope_ids = epi_segments_uniq_res,
  metap_method = "wilkinsons_max1"
)
prot_padj_uniq <- calcProteinPValues(
  epitope_ds = epi_padj_uniq,
  metap_method = "tippetts"
)
prot_calls <- makeProteinCalls(prot_padj_uniq)
```

min_max *Cap vector at minimum/maximum values*

Description

Cap vector at minimum/maximum values

Usage

```
min_max(val, min.value, max.value)
```

Arguments

val vector of values to cap
min.value minimum value
max.value maximum value

Value

vector of capped values

Examples

```
min_max(10, 1, 5)
```

oneHitEpitopes *Find One-hit epitopes*

Description

Find One-hit epitopes

Usage

```
oneHitEpitopes(sample_epitopes)
```

Arguments

sample_epitopes
 logical epitope matrix from makeCalls

Value

vector of one-hit, one-probe epitopes

Examples

```
hit_mat = data.frame(  
  row.names = c("A_1_1", "A_2_2", "A_3_3", "A_4_4"),  
  sample1 = c(TRUE, FALSE, FALSE, TRUE),  
  sample2 = c(TRUE, TRUE, FALSE, FALSE),  
  sample3 = c(TRUE, TRUE, FALSE, FALSE)  
)  
oneHitEpitopes(hit_mat)
```

oneHitProbes *Find one hit probes*

Description

Find one hit probes

Usage

```
oneHitProbes(sample_probes)
```

Arguments

sample_probes logical probe matrix from makeCalls

Value

vector of probes that are one-hits

Examples

```
hit_mat <- data.frame(
  row.names = c("A;1", "A;2", "A;3", "A;4"),
  sample1 = c(TRUE, FALSE, FALSE, TRUE),
  sample2 = c(TRUE, TRUE, FALSE, FALSE),
  sample3 = c(TRUE, TRUE, FALSE, FALSE)
)
oneHitProbes(hit_mat)
```

oneProbeEpitopes *Indicate which epitopes are just one probe.*

Description

Indicate which epitopes are just one probe.

Usage

```
oneProbeEpitopes(epitope_ids)
```

Arguments

epitope_ids vector of epitope ids

Value

vector of logical indicating epitopes that are one probe

Examples

```
oneProbeEpitopes(c("A_1_1", "B_1_1", "C_1_2"))
```

probeHitSupported	<i>Find probe hits with a consecutive probe or another sample</i>
-------------------	---

Description

Find probe hits with a consecutive probe or another sample

Usage

```
probeHitSupported(hit_mat)
```

Arguments

hit_mat matrix of logical values that indicate a hit with a TRUE value

Value

matrix of logical values indicate that the TRUE hit is supported by a consecutive probe hit in the sample sample or the within another sample

pvalue_to_zscore	<i>Convert p-value matrix to a z-score matrix</i>
------------------	---

Description

Convert p-value matrix to a z-score matrix

Usage

```
pvalue_to_zscore(mat.in, one.sided = TRUE, log.p = FALSE, inf.zscore = 16)
```

Arguments

mat.in matrix of p-values
one.sided p-values one-sided
log.p are p-values log transformed?
inf.zscore infinite z-scores are capped to this value

Value

matrix of z-scores

Examples

```
mat <- matrix(runif(100), nrow=10)  
rownames(mat) <- paste0("A;", seq_len(nrow(mat)))  
pvalue_to_zscore(mat)
```

quantileNormalize	<i>Normalize the exprs assay using quantile normalization</i>
-------------------	---

Description

Normalize the exprs assay using quantile normalization

Usage

```
quantileNormalize(se)
```

Arguments

se SummarizedExperiment with exprs assay

Value

SummarizedExperiment with exprs assay normalized

Examples

```
data(heffron2021_wuhan)
seq_ds_qn <- quantileNormalize(heffron2021_wuhan)
```

smoothProbeDS	<i>Smooth probes across protein tiling</i>
---------------	--

Description

Smooth probes across protein tiling

Usage

```
smoothProbeDS(probe_ds, w = 2, eps = 1e-06)
```

Arguments

probe_ds HERONProbeDataSet to smooth
w smoothing width, probes +/- w/2 before and after are used
eps error tolerance

Value

HERONProbeDataSet with smoothed data in exprs object

Examples

```
data(heffron2021_wuhan)
probe_ds <- convertSequenceDSToProbeDS(heffron2021_wuhan)
smoothed_ds <- smoothProbeDS(probe_ds)
```

Index

- * **datasets**
 - heffron2021_wuhan, [17](#)
- * **internal**
 - HERON-package, [3](#)
 - .HERONEpitopeDataSet
 - (HERONEpitopeDataSet-class), [17](#)
 - .HERONProbeDataSet
 - (HERONProbeDataSet-class), [18](#)
 - .HERONProteinDataSet
 - (HERONProteinDataSet-class), [18](#)
 - .HERONSequenceDataSet
 - (HERONSequenceDataSet-class), [19](#)
- addSequenceAnnotations, [3](#)
- calcCombPValues, [4](#)
- calcEpitopePValues, [5](#)
- calcProbePValuesTPaired, [6](#)
- calcProbePValuesTUnpaired, [7](#)
- calcProbePValuesWUnpaired, [7](#)
- calcProteinPValues, [8](#)
- catSequences, [9](#)
- convertSequenceDSToProbeDS, [9](#)
- findBlocksProbeT, [10](#)
- findBlocksT, [10](#)
- findEpitopeSegments, [11](#)
- getEpitopeID, [12](#)
- getEpitopeIDsToProbeIDs, [12](#)
- getEpitopeProbeIDs, [13](#)
- getEpitopeProtein, [13](#)
- getEpitopeStart, [14](#)
- getEpitopeStop, [14](#)
- getKofN, [15](#)
- getProteinLabel, [15](#)
- getProteinStart, [16](#)
- getProteinTiling, [16](#)
- heffron2021_wuhan, [17](#)
- HERON (HERON-package), [3](#)
- HERON-package, [3](#)
- HERONEpitopeDataSet
 - (HERONEpitopeDataSet-class), [17](#)
- HERONEpitopeDataSet-class, [17](#)
- HERONProbeDataSet
 - (HERONProbeDataSet-class), [18](#)
- HERONProbeDataSet-class, [18](#)
- HERONProteinDataSet
 - (HERONProteinDataSet-class), [18](#)
- HERONProteinDataSet-class, [18](#)
- HERONSequenceDataSet
 - (HERONSequenceDataSet-class), [19](#)
- HERONSequenceDataSet-class, [19](#)
- log2Transform, [20](#)
- makeEpitopeCalls, [20](#)
- makeProbeCalls, [21](#)
- makeProteinCalls, [22](#)
- min_max, [22](#)
- oneHitEpitopes, [23](#)
- oneHitProbes, [24](#)
- oneProbeEpitopes, [24](#)
- probeHitSupported, [25](#)
- pvalue_to_zscore, [25](#)
- quantileNormalize, [26](#)
- smoothProbeDS, [26](#)