

# Package ‘Rfastp’

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**Type** Package

**Title** An Ultra-Fast and All-in-One Fastq Preprocessor (Quality Control, Adapter, low quality and polyX trimming) and UMI Sequence Parsing).

**Version** 1.20.3

**Description** Rfastp is an R wrapper of fastp developed in c++. fastp performs quality control for fastq files. including low quality bases trimming, polyX trimming, adapter auto-detection and trimming, paired-end reads merging, UMI sequence/id handling. Rfastp can concatenate multiple files into one file (like shell command cat) and accept multiple files as input.

**License** GPL-3 + file LICENSE

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.1.1

**biocViews** QualityControl, Sequencing, Preprocessing, Software

**SystemRequirements** GNU make

**LinkingTo** Rcpp, Rhtslib

**Imports** Rcpp, rjson, ggplot2, reshape2

**Suggests** BiocStyle, testthat, knitr, rmarkdown

**VignetteBuilder** knitr

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|          |                                 |
|----------|---------------------------------|
| catfastq | <i>Concatenate Fastq Files.</i> |
|----------|---------------------------------|

---

### Description

concatenate multiple fastq files into a single file.

### Usage

```
catfastq(output, inputFiles, append = FALSE, paired = FALSE, shuffled = FALSE)
```

### Arguments

|            |  |
|------------|--|
| output     | output file name [string]  |
| inputFiles | a vector of input file names [vector]  |
| append     | a logical indicating append the files to a file already exists.  |
| paired     | a logical indicating split the input files into two halves. the first half merged into read1, the second half merged into read2. |
| shuffled   | a logical indicating split the input file list into two halves. The R1/R2 files are interleaved in the inputFiles vector.        |

### Value

no returns.

### Author(s)

Wei Wang

### Examples

```
pe001_read1 <- system.file("extdata", "splited_001_R1.fastq.gz",
  package="Rfastp")
pe002_read1 <- system.file("extdata", "splited_002_R1.fastq.gz",
  package="Rfastp")
pe003_read1 <- system.file("extdata", "splited_003_R1.fastq.gz",
  package="Rfastp")
pe004_read1 <- system.file("extdata", "splited_004_R1.fastq.gz",
  package="Rfastp")

pe001_read2 <- system.file("extdata", "splited_001_R2.fastq.gz",
  package="Rfastp")
pe002_read2 <- system.file("extdata", "splited_002_R2.fastq.gz",
```

```

    package="Rfastp")
pe003_read2 <- system.file("extdata","splited_003_R2.fastq.gz",
    package="Rfastp")
pe004_read2 <- system.file("extdata","splited_004_R2.fastq.gz",
    package="Rfastp")

allR1 <- c(pe001_read1, pe002_read1, pe003_read1, pe004_read1)
allR2 <- c(pe001_read2, pe002_read2, pe003_read2, pe004_read2)

allreads <- c(allR1, allR2)
allreads_shuffled <- c(pe001_read1, pe001_read2, pe002_read1, pe002_read2,
    pe003_read1, pe003_read2, pe004_read1, pe004_read2)

outputPrefix <- tempfile(tmpdir = tmpdir())
# a normal concatenation for single-end libraries.

catfastq(output = paste0(outputPrefix, "_R1.fastq.gz"), inputFiles = allR1)

# a normal concatenation for paired-end libraries.

catfastq(output = paste0(outputPrefix, "merged_paired"),
    inputFiles = allreads, paired=TRUE)

# Append to exist files (paired-end)

catfastq(output=paste0(outputPrefix,"append_paired"), inputFiles=allreads,
    append=TRUE, paired=TRUE)

# Input paired-end files are shuffled.

catfastq(output=paste0(outputPrefix,"_shuffled_paired"),
    inputFiles=allreads_shuffled, paired=TRUE, shuffled=TRUE)

```

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curvePlot

*Plot of Base Quality and GC Content.*


---

### Description

generate a ggplot2 object of Base Quality/GC content before and after QC.

### Usage

```
curvePlot(json, curves = "quality_curves")
```

### Arguments

|        |   |
|--------|---|
| json   | the output json of function rfastq. [json]  |
| curves | plots for Base Quality("quality_curves") or GC content("content_curves"). default is "quality_curves" |

### Value

a ggplot2 object.

**Author(s)**

Wei Wang

**Examples**

```
outputPrefix <- tempfile(tmpdir = tempdir())
se_read1 <- system.file("extdata", "Fox3_Std_small.fq.gz", package="Rfastp")
se_json_report <- rfastp(read1 = se_read1, outputFastq = outputPrefix,
  thread = 4)
# Base Quality plot is the default output:
p1 <- curvePlot(se_json_report)
p1

p2 <- curvePlot(se_json_report, curves = "content_curves")
```

---

qcSummary

*Summary of Fastq Quality Control*

---

**Description**

generate a data frame of the Fastq QC summary.

**Usage**

```
qcSummary(json)
```

**Arguments**

json                    the output json of function rfastq. [json]

**Value**

a data frame.

**Author(s)**

Wei Wang

**Examples**

```
outputPrefix <- tempfile(tmpdir = tempdir())
se_read1 <- system.file("extdata", "Fox3_Std_small.fq.gz", package="Rfastp")
se_json_report <- rfastp(read1 = se_read1, outputFastq = outputPrefix,
  thread = 4)
df_summary <- qcSummary(se_json_report)
```

---

`rfastp`*R wrap of fastp*

---

**Description**

Quality control (Cut adapter, low quality trimming, polyX trimming, UMI handling, and etc.) of fastq files.

**Usage**

```
rfastp(  
  read1,  
  read2 = "",  
  outputFastq,  
  unpaired = "",  
  failedOut = "",  
  merge = FALSE,  
  mergeOut = "",  
  phred64 = FALSE,  
  interleaved = FALSE,  
  fixMGId = FALSE,  
  adapterTrimming = TRUE,  
  adapterSequenceRead1 = "auto",  
  adapterSequenceRead2 = "auto",  
  adapterFasta = "",  
  trimFrontRead1 = 0,  
  trimTailRead1 = 0,  
  trimFrontRead2 = 0,  
  trimTailRead2 = 0,  
  maxLengthRead1 = 0,  
  maxLengthRead2 = 0,  
  forceTrimPolyG = FALSE,  
  disableTrimPolyG = FALSE,  
  minLengthPolyG = 10,  
  trimPolyX = FALSE,  
  minLengthPolyX = 10,  
  cutWindowSize = 4,  
  cutLowQualTail = FALSE,  
  cutSlideWindowRight = FALSE,  
  cutLowQualFront = FALSE,  
  cutMeanQual = 20,  
  cutFrontWindowSize = 4,  
  cutFrontMeanQual = 20,  
  cutTailWindowSize = 4,  
  cutTailMeanQual = 20,  
  cutSlideWindowSize = 4,  
  cutSlideWindowQual = 20,  
  qualityFiltering = TRUE,  
  qualityFilterPhred = 15,  
  qualityFilterPercent = 40,  
  maxNfilter = 5,  
)
```

```

averageQualFilter = 0,
lengthFiltering = TRUE,
minReadLength = 15,
maxReadLength = 0,
lowComplexityFiltering = FALSE,
minComplexity = 30,
index1Filter = "",
index2Filter = "",
maxIndexMismatch = 0,
correctionOverlap = FALSE,
minOverlapLength = 30,
maxOverlapMismatch = 5,
maxOverlapMismatchPercentage = 20,
umi = FALSE,
umiLoc = "",
umiLength = 0,
umiPrefix = "",
umiSkipBaseLength = 0,
umiNoConnection = FALSE,
umiIgnoreSeqNameSpace = FALSE,
overrepresentationAnalysis = FALSE,
overrepresentationSampling = 20,
splitOutput = 0,
splitByLines = 0,
thread = 2,
verbose = TRUE
)

```

### Arguments

|                      |   |
|----------------------|---|
| read1                | read1 input file name(s). [vector]  |
| read2                | read2 input file name(s). [vector]  |
| outputFastq          | string of /path/prefix for output fastq [string]  |
| unpaired             | for PE input, output file name for reads which the mate reads failed to pass the QC [string], default NULL, discard it. [string]  |
| failedOut            | file to store reads that cannot pass the filters default NULL, discard it. [string]   |
| merge                | for PE input, A logical(1) indicating whether merge each pair of reads into a single read if they are overlaped, unmerged reads will be write to 'output' file. Default is FALSE. the 'mergeOut' must be set if TRUE. |
| mergeOut             | under 'merge' mode, file to store the merged reads. [string]  |
| phred64              | A logical indicating whether the input is using phred64 scoring (it will be converted to phred33, so the output will still be . phred33)  |
| interleaved          | A logical indicating whether <read1> is an interleaved FASTQ which contains both read1 and read2. Default is FALSE.   |
| fixMGIIid            | the MGI FASTQ ID format is not compatible with many BAM operation tools, enable this option to fix it. Default is FALSE   |
| adapterTrimming      | A logical indicating whether run adapter trimming. Default is 'TRUE'  |
| adapterSequenceRead1 | the adapter for read1. For SE data, if not specified, the adapter will be auto-detected. For PE data, this is used if R1/R2 are found not overlapped.   |

|                      |   |
|----------------------|---|
| adapterSequenceRead2 | the adapter for read2 (PE data only). This is used if R1/R2 are found not overlapped. If not specified, it will be the same as <adapterSequenceRead1>   |
| adapterFasta         | specify a FASTA file to trim both read1 and read2 (if PE) by all the sequences in this FASTA file.  |
| trimFrontRead1       | trimming how many bases in front for read1, default is 0.   |
| trimTailRead1        | trimming how many bases in tail for read1, default is 0'  |
| trimFrontRead2       | trimming how many bases in front for read2. If it's not specified, it will follow read1's settings  |
| trimTailRead2        | trimming how many bases in tail for read2. If it's not specified, it will follow read1's settings   |
| maxLengthRead1       | if read1 is longer than maxLengthRead1, then trim read1 at its tail to make it as long as maxLengthRead1. Default 0 means no limitation.  |
| maxLengthRead2       | if read2 is longer than maxLengthRead2, then trim read2 at its tail to make it as long as maxLengthRead2. Default 0 means no limitation. If it's not specified, it will follow read1's settings.    |
| forceTrimPolyG       | A logical indicating force polyG tail trimming, trimming is only automatically enabled for Illumina NextSeq/NovaSeq . data.   |
| disableTrimPolyG     | A logical indicating disable polyG tail trimming.   |
| minLengthPolyG       | the minimum length to detect polyG in the read tail. 10 by default.   |
| trimPolyX            | A logical indicating force polyX tail trimming.   |
| minLengthPolyX       | the minimum length to detect polyX in the read tail. 10 by default.   |
| cutWindowSize        | the window size option shared by cutLowQualFront, cutLowQualTail, or cutSlideWindowRight. Range: 1~1000, default: 4   |
| cutLowQualTail       | A logical indiccating move a sliding window from tail (3') to front, drop the bases in the window if its mean quality < threshold, stop otherwise. Default is 'FALSE'                               |
| cutSlideWindowRight  | A logical indicating move a sliding window from front to tail, if meet one window with mean quality < threshold, drop the bases in the window and the right part, and then stop. Default is 'FALSE' |
| cutLowQualFront      | A logical indiccating move a sliding window from front (5') to tail, drop the bases in the window if its mean quality < threshold, stop otherwise. Default is 'FALSE'                               |
| cutMeanQual          | the mean quality requirement option shared by cutLowQualFront, cutLowQualTail or cutSlideWindowRight. Range: 1~36, default: 20  |
| cutFrontWindowSize   | the window size option of cutLowQualFront, default to cutWindowSize if not specified. default: 4  |
| cutFrontMeanQual     | the mean quality requirement option for cutLowQualFront, default to cutMeanQual if not specified. default: 20   |
| cutTailWindowSize    | the window size option of cutLowQualTail, default to cutWindowSize if not specified. default: 4   |

|                        |   |
|------------------------|---|
| cutTailMeanQual        | the mean quality requirement option for cutLowQualTail, default to cutMeanQual if not specified. default: 20  |
| cutSlideWindowSize     | the window size option of cutSlideWindowRight, default to cutWindowSize if not specified. default: 4  |
| cutSlideWindowQual     | the mean quality requirement option for cutSlideWindowRight, default to cutMeanQual if not specified. default: 20   |
| qualityFiltering       | A logical indicating run quality filtering. Default is 'TRUE'.  |
| qualityFilterPhred     | the minimum quality value that a base is qualified. Default 15 means phred quality $\geq Q15$ is qualified.   |
| qualityFilterPercent   | Maximum percents of bases are allowed to be unqualified (0~100). Default 40 means 40%   |
| maxNfilter             | maximum number of N allowed in the sequence. read/pair is discarded if failed to pass this filter. Default is 5   |
| averageQualFilter      | if one read's average quality score $<$ 'averageQualFilter', then this read/pair is discarded. Default 0 means no requirement.  |
| lengthFiltering        | A logical indicating whether run length filtering. Default: TRUE  |
| minReadLength          | reads shorter than minReadLength will be discarded, default is 15.  |
| maxReadLength          | reads longer than maxReadLength will be discarded, default 0 means no limitation.   |
| lowComplexityFiltering | A logical indicating whether run low complexity filter. The complexity is defined as the percentage of base that is different from its next base ( $\text{base}[i] \neq \text{base}[i+1]$ ). Default is 'FALSE' |
| minComplexity          | the threshold for low complexity filter (0~100). Default is 30, which means 30% complexity is required. (int [=30])   |
| index1Filter           | specify a file contains a list of barcodes of index1 to be filtered out, one barcode per line.  |
| index2Filter           | specify a file contains a list of barcodes of index2 to be filtered out, one barcode per line.  |
| maxIndexMismatch       | the allowed difference of index barcode for index filtering, default 0 means completely identical.  |
| correctionOverlap      | A logical indicating run base correction in overlapped regions (only for PE data), default is 'FALSE'   |
| minOverlapLength       | the minimum length to detect overlapped region of PE reads. This will affect overlap analysis based PE merge, adapter trimming and correction. 30 by default.   |

|                              |   |
|------------------------------|---|
| maxOverlapMismatch           | the maximum number of mismatched bases to detect overlapped region of PE reads. This will affect overlap analysis based PE merge, adapter trimming and correction. 5 by default.            |
| maxOverlapMismatchPercentage | the maximum percentage of mismatched bases to detect overlapped region of PE reads. This will affect overlap analysis based PE merge, adapter trimming and correction. Default 20 means 20% |
| umi                          | A logical indicating whether preprocessing unique molecular identifier (UMI). Default: 'FALSE'  |
| umiLoc                       | specify the location of UMI, can be (index1/index2/read1/read2/per_index/per_read)  |
| umiLength                    | length of UMI if the UMI is in read1/read2.   |
| umiPrefix                    | an string indication the following string is UMI (i.e. prefix=UMI, UMI=AATTCG, final=UMIAATTCG). Only letters, numbers, and '#' allowed. No prefix by default.                              |
| umiSkipBaseLength            | if the UMI is in read1/read2, skip 'umiSkipBaseLength' bases following UMI, default is 0.   |
| umiNoConnection              | an logical indicating remove "_" between the UMI prefix string and the UMI string. Default is FALSE.  |
| umiIgnoreSeqNameSpace        | an logical indicating ignore the space in the sequence name. Default is FALSE, the umi tag will be inserted into the sequence name before the first SPACE.                                  |
| overrepresentationAnalysis   | A logical indicating overrepresentation analysis. Default is 'FALSE'  |
| overrepresentationSampling   | one in 'overrepresentationSampling' reads will be computed for overrepresentation analysis (1~10000), smaller is slower, default is 20.   |
| splitOutput                  | number of files to be splitted (2~999). a sequential number prefix will be added to output name. Default is 0 (no split)  |
| splitByLines                 | split output by limiting lines of each file(>=1000), a sequential number prefix will be added to output name ( 0001.out.fq, 0002.out.fq...), default is 0 (disabled).                       |
| thread                       | owrker thread number, default is 2  |
| verbose                      | output verbose log information  |

**Value**

returns a json object of the report.

**Author(s)**

Thomas Carroll, Wei Wang

**Examples**

```
# prepare for the input and output files.
# if the output file exists, it will be OVERWRITEN.
```

```
se_read1 <- system.file("extdata", "Fox3_Std_small.fq.gz", package="Rfastp")
pe_read1 <- system.file("extdata", "reads1.fastq.gz", package="Rfastp")
pe_read2 <- system.file("extdata", "reads2.fastq.gz", package="Rfastp")
outputPrefix <- tempfile(tmpdir = tmpdir())

# a normal single-end file

se_json_report <- rfastp(read1 = se_read1,
  outputFastq=paste0(outputPrefix, "_se"), thread = 4)

# merge paired-end data by overlap:

pe_json_report <- rfastp(read1 = pe_read1, read2 = pe_read2, merge = TRUE,
  outputFastq = paste0(outputPrefix, '_unpaired'),
  mergeOut = paste0(outputPrefix, '_merged.fastq.gz'))

# a clipr example

clipr_json_report <- rfastp(read1 = se_read1,
  outputFastq = paste0(outputPrefix, '_clipr'),
  disableTrimPolyG = TRUE,
  cutLowQualFront = TRUE,
  cutFrontWindowSize = 29,
  cutFrontMeanQual = 20,
  cutLowQualTail = TRUE,
  cutTailWindowSize = 1,
  cutTailMeanQual = 5,
  minReadLength = 29,
  adapterSequenceRead1 = 'GTGTCAGTCACTTCCAGCGG'
)
```

---

trimSummary

*Summary of Fastq adapter and low quality trimming*

---

## Description

generate a data frame of the Fastq trim summary.

## Usage

```
trimSummary(json)
```

## Arguments

json                    the output json of function rfastq. [json]

## Value

a data frame.

**Author(s)**

Wei Wang

**Examples**

```
outputPrefix <- tempfile(tmpdir = tmpdir())
se_read1 <- system.file("extdata", "Fox3_Std_small.fq.gz", package="Rfastp")
se_json_report <- rfastp(read1 = se_read1, outputFastq = outputPrefix,
  thread = 4, adapterSequenceRead1 = 'GTGTCAGTCACTTCCAGCGG')
trim_summary <- trimSummary(se_json_report)
```

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